# The Complete Mitochondrial DNA Sequence of the Bichir (Polypterus ornatipinmis), a Basal Ray-Finned Fish: Ancient Establishment of the Consensus Vertebrate Gene Order 

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#### Abstract

The evolutionary position of bichirs is disputed, and they have been variously aligned with ray-finned fish (Actinopterygii) or lobe-finned fish (Sarcopterygii), which also include tetrapods. Alternatively, they have been placed into their own group, the Brachiopterygii. The phylogenetic position of bichirs as possibly the most primitive living bony fish (Osteichthyes) made knowledge about their mitochondrial genome of considerable evolutionary interest. We determined the complete nucleotide sequence ( 16,624 bp ) of the mitochondrial genome of a bichir, Polypterus omatipinnis. Its genome contains 13 proteincoding genes, 22 tRNAs, two rRNAs and one major noncoding region. The genome's structure and organization show that this is the most basal vertebrate that conforms to the consensus vertebrate mtDNA gene order. Bichir mitochondrial protein-coding and ribosomal RNA genes have greater sequence similarity to ray-finned fish than to either lamprey or lungfish. Phylogenetic analyses suggest the bichir's placement as the most basal living member of the ray-finned fish and rule out its classification as a lobefinned fish. Hence, its lobe-fins are probably not a shared-derived trait with those of lobe-finned fish (Sarcopterygii).


BONY fish (Osteichthyes) are typically divided into two major groups, the actinopterygians (rayfinned fish) and the sarcopterygians (lobe-finned fish). With $>25,000$ known species, the ray-finned fish comprise by far the largest class of vertebrates. Ray-finned fish date back to the early Devonian (Carroll 1988), and their diversity has since then increased steadily both in terms of morphological diversity and in terms of numbers of species. By contrast, the once highly successful and diverse lobe-finned fish, now only include two extant fish groups, the lungfish and the coelacanth. During the last 150 yr , both of these latter groups have been variously thought to be the closest living relatives of land vertebrates (e.g., Zardoya and Meyer 1996a; reviewed in Meyer 1995).

While the classification of bony fish into either the ray-finned or the lobe-finned fish is without problems, the phylogenetic placement of species of the order Polypteriformes into one of these two categories (e.g., Patterson 1982; Lauder and Liem 1983, also referred to as Cladistia) has been historically difficult, and it still remains somewhat uncertain (Table 1). Unlike any other group of living fish (reviewed in Patterson 1982; Lauder and Liem 1983), polypteriform fish (bichirs, Polypterus, and reedfish, Calamoichthys) have been placed into a number of widely differing taxonomic groups during the last century.

[^0]Figure 1 summarizes some of the various phylogenetic hypotheses involving bichirs. One of the earliest hypotheses of polypterid ancestry held that they were sarcopterygians, and bichirs were placed into the suborder Crossopterygii along with fossil rhipidistians, coelacanths and lungfish (Figure la) (Huxley 1861; COPE 1871; reviewed in Patterson 1980, 1982). Goodrich (1928), for the first time, considered placing polypterids within the actinopterygians as a taxonomically distinct lower division. Since then, the alternative two main hypotheses of polypterine relationships have been that polypterids are either classified as a distinct subclass of Osteichthyes, the Brachiopterygii (Figure 1b), or as basal actinopterygians (Figure 1c). The main problems of assessing polypterine relationships are that the fossil record for these fish only extends back to the Eocene (Greenwood 1974; Carroll 1988) and that they show a large number of primitive ancestral as well as uniquely derived characteristics that make the determination of their phylogenetic position, and hence their classification, difficult. Bichirs have a row of dorsal finlets that extend down their back, primitive rhombic ganoid scales, numerous spiracular plates, and lobed pectoral fins. Their pectoral fins have a fleshy external appearance similar to those of lungfish and the coelacanth. Polypterine fish also have a well-vascularized lung that arises from the ventral foregut and a pair of external gills in juveniles, a rare feature, not seen in any other fish, except in some species of lungfish.

Many authors still argue that Polypteriformes are

TABLE 1

## Systematic position of bichirs

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Superclass: Gnathostomata
    Class Chondrichthyes
        Subclass: Holocephali (Chimaeras)
        Subclass: Elasmobranchii (sharks, rays)
    Class Sarcopterygii
        Subclass: Coelacanthimorpha (Latimeria)
        Subclass: Dipnoi (lungfishes)
        Subclass: Tetrapoda (amphibians, reptiles, birds,
                mammals)
    Class Actinopterygii
        Subclass: Chondrostei
            Order: Polypteriformes (bichirs, Polypterus,
                reedfishes, Calamoichthys)
            Order: Acipenseriformes (sturgeons, Acipenser;
                paddlefishes, Polyodont)
        Subclass: Neopterygii
            Order: Semionotiformes (gars, Lepisosteus)
            Order: Amiiformes (bowfins, Amia)
                Division: Teleostei
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    Modified from Nelson (1994)
    A


Bichirs

Lobe-finned fish

Tetrapods

Ray-finned fish
morphologically too distinct from actinopterygians to be classified as such (Jessen 1973; Nelson 1973; Jarvik 1980; Bjerring 1985). This view, however, does not clarify what other group of fish polypterids are most closely related to. Most recent authors have, based on morphological characters such as scale structure and neurocranial ossification, placed polypterids with actinopterygians (Figure 1c) (Gardiner 1973; Schaeffer 1973; Patterson 1980, 1982; Lauder and Liem 1983; GardiNer and schaeffer 1989). However, the placement of Polypteriformes within the Actinopterygii is also uncertain. Often polypterids are placed within the Chondrostei, a group to which most authors also add the sturgeons and their relatives (Table 1, e.g., Gardiner 1967; Andrews 1967; Carroll 1988; Nelson 1994). Whereas other researchers classify polypterids as a taxon of equal rank and basal to the Chondrostei (reviewed in Patterson 1982; Lauder and Liem 1983).

Among other reasons, knowledge of the phylogenetic position of the bichir is of importance for the reconstruction of the development and evolution of verte-

B


Bichirs
Ray-finned fish
JARVIK 1980 BJERRING 1985

Tetrapods

C


Bichirs

| Ray-finned fish | GARDINER 1973 <br> SCHAEFFER 1973 |
| :--- | :--- |
| Lobe-finned fish | PATTERSON 1982 <br> LAUDER \& LIEM 1982 <br>  |
| SCtrapods | SCHAEFFER 1989 |
| NELSON 1994 |  |

brate traits such as paired limbs from fins (e.g., SHubin and Alberch 1986; Sordino and Duboule 1996). However, so far, only two molecular phylogenetic studies have incorporated nucleotide sequence data from bichirs. NORMARK et al. (1991) used partial DNA sequences of the mitochondrial cyt $b, C O I$ and COII genes to infer phylogenetic relationships of Neopterygians, using the bichir as outgroup taxon. Lê et al. (1993) used portions of the large nuclear ribosomal gene in an attempt to infer the phylogeny among major fish lineages. Their study supported the view that polypterids are a primitive ray-finned fish lineage, albeit with low confidence.

Mitochondrial genes are increasingly used to infer phylogenetic relationships both among closely related species as well as distantly related ones (Kumazawa and Nishida 1993; CaO et al. 1994; Russo et al. 1996; Zardoya and Meyer 1996c). Zardoya and Meyer (1996a) show that combined sets of all mitochondrial protein coding and tRNA genes can with strong support resolve the relationships among major groups of jawed vertebrates. Until now the complete mitochondrial genome sequences of 22 vertebrate species have been reported; 15 are from mammals, five from fish, only one from an amphibian and one from a bird. The structure and organization of vertebrate mitochondrial genomes is remarkably conserved. Minor rearrangements, apparently caused by the translocation of tRNA genes, have been described for chicken (Desjardins and Morais 1990) and for the opposum (Janke et al. 1994). It is not yet clear when the establishment of the vertebrate consensus gene order occurred during their evolution. Lampreys, one of the earliest vertebrates, have a peculiar gene order (Lee and Kocher 1995), but without the study of more chordates, it is impossible to determine if this gene order represents an ancestral or a uniquely derived condition. We determined the complete nucleotide sequence and gene order of the bichir mitochondrial genome with the aim to investigate the evolution of the vertebrate mitochondrial genome and to clarify the bichir's relationships to lobe-finned and ray-finned fish. Furthermore, we assessed the performance of the bichir as an outgroup taxon for inferring relationships among major groups of jawed vertebrates.

## MATERIALS AND METHODS

[^1]3.1-kb fragment was amplified using specific oligonucleotide primers and then cloned into pGEM-T (Promega).
Plasmid DNA was extracted from each clone using a Wizard miniprep kit (Promega). After ethanol precipitation, purified DNA was used as template for Taq Dye Deoxy Terminator cycle-sequencing reactions (Applied Biosystems Inc.) following manufacturer's instructions. Sequencing was performed with an automated DNA sequencer (373A Stretch, Applied Biosystems Inc.). Sequences were obtained using both M13 universal sequencing primers and 37 specifically designed oligonucleotide primers. The sequences obtained from each clone were $\sim 350 \mathrm{bp}$ in length and each sequence overlapped the next by $\sim 100 \mathrm{bp}$. In no case were differences in sequence observed among the overlapping regions.

Sequence data were analyzed by use of the GCG program package (Devereux et al. 1984) and alignments were performed using CLUSTAL W (Thompson et al. 1994). The data were subjected to maximum parsimony (MP) analyses using PAUP Version 3.1.1 (Swofford 1993). MP analyses were performed using heuristic searches (TBR branch swapping, MULPARS option in effect, simple stepwise addition of taxa). Maximum likelihood (ML) analyses and neighbor-joining (NJ) analyses were performed using PHYLIP Version 3.5 (Felsenstein 1989). For NJ analyses, distance matrices were calculated based on Kimura corrected distances (Kimura 1980). The Kishino-Hasegawa test to assess the statistical confidence of maximum likelihood trees was performed using MOLPHY Version 2.2 (Adachi and Hasegawa 1992). Confidence levels for MP and NJ analyses were assessed by bootstrap analyses based on 1000 replications (Felsenstein 1985) using PHYLIP Version 3.5 (Felsenstein 1989).
The complete mtDNA sequence of the bichir has been deposited in GenBank under the accession No. U62532.

## RESULTS AND DISCUSSION

Genome organization: The total length of the bichir mitochondrial genome is $16,624 \mathrm{bp}$, it is similar to that of the mitochondrial genomes of other bony fish (Table 2). The overall base composition of the L-strand is A $=32.2 \%, \mathrm{~T}=28.1 \%, \mathrm{C}=25.6 \%$ and $\mathrm{G}=14.1 \%$. As found in other vertebrates, the bichir's mitochondrial genome contains 13 protein coding genes, 22 transfer RNA genes, and two ribosomal RNA genes. Interestingly, the orientation and relative position of all genes and the major noncoding region is identical to the vertebrate consensus mitochondrial gene order (Figures 2 and 3, Table 3). The presence of this gene order in the bichir documents the early establishment of the particular vertebrate organization of the mitochondrial genome. All genes except the gene encoding for ND6 and eight transfer RNAs are encoded on the H-strand, and the genes encoded on both strands of the bichir mtDNA are of similar length as those in other vertebrates (Table 3). Peptide encoding genes were identified by comparison with the rainbow trout mtDNA (Zardoya et al. 1995) and by the presence of initiation and stop codons. Transfer RNA genes were identified by sequence similarity to homologous vertebrate tRNAs, their specific anticodons, and their ability to fold into putative cloverleaf structures.

As seen in other vertebrates there is an overlap between the genes encoding for ND4 and ND4L, ATPase

TABLE 2
Lengths of fish mitochondrial genes

|  | Species |  |  |  |  |  |
| :--- | :---: | ---: | ---: | ---: | ---: | ---: |
| Gene | Lungfish | Trout | Carp | Loach | Bichir | Lamprey |
| Control region | 1184 | 1003 | 927 | 896 | 1068 | 491 |
| 12S rRNA | 937 | 944 | 951 | 937 | 950 | 900 |
| I6S rRNA | 1591 | 1680 | 1681 | 1680 | 1655 | 1621 |
| Cytb | 1144 | 1141 | 1141 | 1144 | 1141 | 1191 |
| ND1 | 966 | 972 | 975 | 975 | 958 | 966 |
| ND2 | 1028 | 1050 | 1047 | 1047 | 1036 | 1044 |
| ND3 | 346 | 349 | 351 | 351 | 346 | 351 |
| ND4 | 1384 | 1381 | 1383 | 1383 | 1378 | 1377 |
| ND4L | 297 | 297 | 297 | 297 | 297 | 291 |
| ND5 | 1836 | 1839 | 1894 | 1837 | 1842 | 1797 |
| ND6 | 513 | 522 | 519 | 522 | 504 | 519 |
| COI | 1548 | 1551 | 1551 | 1551 | 1557 | 1554 |
| COII | 691 | 691 | 691 | 691 | 688 | 690 |
| COIII | 784 | 784 | 786 | 768 | 784 | 786 |
| APT6 | 682 | 670 | 684 | 684 | 682 | 714 |
| ATP8 | 168 | 168 | 165 | 168 | 168 | 168 |
| Total | 16646 | 16642 | 16575 | 16558 | 16624 | 16201 |

Values expressed as base pairs.

6 and ATPase 8, and ND5 and ND6. The bichir mtDNA also has an overlap of nine nucleotides involving COI and $t R N A^{\operatorname{Ser}(U C N)}$, which are encoded on opposite DNA strands. This overlap is also found in the lamprey (LEE and Kocher 1995) but not in any other fish (Tzeng et al. 1992; Chang et al. 1994; Zardoya et al. 1995; ZARdoya and Meyer 1996a) where these two genes either directly abutt or are separated by a few nucleotides. These data show that the birchir is the most basal vertebrate known so far with the consensus vertebrate mito-
chondrial gene order. Once a complete chondrichthyan mitochondrial genome has been sequenced, the exact phylogenetic timing of the evolution of this gene order in vertebrates will have been established.

Protein coding genes: The bichir mitochondrial genome encodes for 13 proteins. Pairwise comparisons of the protein coding genes of the bichir with their homologues of rainbow trout, loach, carp, lungfish and lamprey showed that bichir proteins consistently have greater sequence similarity to those from ray-finned fish


Figure 2.-Restriction map and gene organization of the Polypterus ornatipinnis mitochondrial genome. All protein coding genes are encoded by the H -strand with the exception of ND6, which is coded by the L-strand. Each tRNA gene is identified by the single letter amino acid code and depicted according to the coding strand. Only the EcoRI and HindIII restriction sites used for cloning are shown.

GCTAGTGTAGCTTATCCTCAAAGCATAACACTGAAAATGTTAAGACGGACCCTAGAAAAGTCCCACAAGCACAAAGATTTGGTCCTAGTCTTACTATCAT
100

tRNA-Val $\rightarrow$

16 S rRNA -

GGAAGGTGCACTTGGAATAACEAAAATGTAGCTTAATAGAAAAGTATCTCCCTTACACTGAGATGAAGTCTGCGCAATACACACCATTTTGAGCATTATA 1100 GCTAGCCTGACCACACACTAAACCAAACACTATTAAAATAAATATACTTATAATAAAAACAAAACATTTGTTAACTTCAGTATAGGCGATAGAAAGAGAA 1200 CAAAAGAGCTATAGCAACAGTACCGCAAGGGAAAGCTGAAAGAGAAATGAAACAAATCGTTAAAGCACGACATAGCAGAGATTAAATCTCGTACCTTTTG CATCATGATCTAGTAAGTAAGCCCAAGCAAAATGATTTATAGTTTGACCCCCCGAAACTAGACGAGCTACTTCGAGGCAGTTGAAAGGACCACCCCGTCT 1400 CTGTGGAAAAAGAGTGGGAAGACTTCCAAGTAGAGGTGACAAGCCTAACGAGCCTAGTGATAGCTGGTTACTTGAGAAATGGATAAAAGTCCAGCCTCAA I5OO AATTTCTAAAAAATATACAAATTCCGTTTAAAAAATTTTAAGAACATTTGAGAGTTATTCACAGGAGGTACAGCTCCTATGAACTGGGAAACAACCCAAT AAGGAGGAAAAAGATCATAATTTACAAGGACAAAATCCAAGTGGGCCTGAAAGCAGCCACCTTTAAAGAAAGCGTTATAGCTTAAATAATATATTATTCC GTATATCCGGATAAAATCTCTGAATCCCCTACAAATATCAAGTTATTCTATTTGAGTAGAAGAAATTATGCTAGAACTAGTAATAAGAAAAATGATTTTC TCCTAGCACAAGTGTAAGTTAGAACGGACAAACCACTAACAATTAGACGAACCCAACAGAGGGCCAAATAACGCCTATAATAAAAACAAGAAAACCCTAT TAATCTTATCGTAAATCTTACACAAGAGTGCCTAAAGGAAAGACTAAAAGAGAAAAAAGGAACTCGGCAAATCCGAGCCCCGCCTGTTTACCAAAAACAT CGCCTTCAGCTTTTCATGTATTGAAGGTCCTGCCTGCCCAGTGACATGAGTTTAACGGCCGCGGTATCCTGACCGTGCAAAGGTAGCGTAATCACTTGTT eTTTAAATGAGGACTGGIATGAATGGCCCCACGAGGGCTCAACTGTCTCTTTTTCTCCGGTCAATTAAACTGATCTCCCCTGTGCAGAAGCGGGCATAAA GACATAAGACGAGAAGACCCTGTGGAGCTTTAGACTAAATCCAAACACTCCTCACTATATTTTACCGTATAGACAAACACAGCGTTATGGCCATAAAGTC TTAGGTTGGGGCGACCACTGAGAACAAATAATCCTCAGCGATGATTGAAGCACAGCTTTATAAACTAAGAATGACAATTCAAAGCATCAGGACACCTGAC ATTAGGATCCAGACTAATCTGATCAACGAACCAAGTTACCCCAGGGATAACAGCGCAATCTTTTCCAAGAGCCCAAATCGACGAAAAGGTTTACGACCTC GATGTTGGATCAGGACATCCTAATGGTGCATCCGCTATTAAGGGTTCGTTTGTTCAGCGATTAAAGTCCTACGTGATCTGAGTTCAGACCGGAGCAATCC AGGTCAGTTTCTATCTATGTAGTCATTTATCCTAGTACGAAAGGATTGGATAAATGGGGCCTATATTAAAGACACGCCCCCTTTTAACCTACTGAAGCCA tRNA-Leu (UUR) $\rightarrow$
AATCAAGTAGATAATAAAAAACATACCCACGCCCTAGAACAGGGTTAGTTAGAGTGGCAGAGCCTGGTAATTGCATGAGACCTAAACCCTCAATCGCAGG NADH1 $\rightarrow$
 GGTTCAACCCCCCTCTITAACTATGACCTTAATCACATTAATTATTAACCCACTCATATATATTATCCCCATTCTGCTAGCAATAGCCTTCCTCACTCTA
 GTAGAACGAAAAATATTAGGCTACATGCAACTCCGAAAAGGACCTAACATCGTAGGCCCATACGGTCTCCTCCAACCCATCGCAGACGGTGTAAAACTAT
 TTATTAAAGAACCAGTAAAACCATCAACCTCTTCCCCAACACTTTTTCTACTCACCCCGACACTTGCCCTCACCTTAGCCCTCATTTTATGAATTCCTCT
 TCCTATACCACTAGCCCTCACAGACCTAAATTTAACTATTCTTTTCATTTTAGCCGTATCAAGCCTATCAGTCTATTCAATCTTAGGTTCAGGTTGAGCT
 TCAAATTCCAAATATGCCCTAATTGGAGCATTACGAGCAGTAGCTCAGACAATTTCATATGAAGTCACCTTAGGCCTCATCATCATCTCCTTAATTATAT
 TTACAGGGGGATTCACATTAACCACCTTCAATACCGCACAAGAAGCCGTATGACTTATCCTACCAGCCTGACCACTAGCCGCAATATGATTCATTTCAAC
 CCGAAACAAACCGAGCACCATTTGATCTTACAGAAGGAGAGTCCGAACTAGTATCTGGATTTAATGTTGAATACGCAGGCGGACCATICGCCCTC


Figure 3.-Complete nucleotide sequence of the L-strand of the bichir mitochondrial DNA molecule. Position 1 refers to the first nucleotide of $t R N A^{\text {Phe }}$. Direction of transcription for each gene is denoted by arrows. The deduced amino acid sequence for each gene product is shown above the nucleotide sequence (one-letter amino acid abbreviation is placed above the first nucleotide of each codon). Termination codons are indicated by an asterisk. tRNA genes are underlined and the corresponding anticodons are overlined. In the control region, CSB (Conserved Sequence Block) and TAS (Termination Associated Sequence) are underlined.
3601 GAGCCATCAAAACAATAATCTTAGCATCCATATTCCTCTGAGTTCGCGCCTCCTACCCTCGATTCCGTTATGATCAACTCATACACTTAGTATGAAAAAA

3701 TTMCCTACOCCTCACICTTGCTTTAATTACCTGACATATTTCACTACCAATCTCCATGGCAGGTTCACCACCGCAACTATAGGCCCTATGCCOGAAAGUT3801 AAGGACNACMTGGTAGAGTAGAAAATAGGGGNCAAACCCCCTTAGTGCCTAGGAABGAAGGACTTGAACCTCCACTAAAAAGATCABAACTTTCAGTG$\leftarrow$ tRNA-GlnNADH $2 \rightarrow$tRNA-Met $\rightarrow$M N $\mathrm{P} \mathbf{Y}$3901 GTHCCATHACACCATCTCCTAGTAAGGTGAGCTGATTAAGCTHTTGGOCCCATACCCCAAATATGHTGGTTAAAATCCITCCCCTACTAATGAACCCATA4001 CATCCTATOCATCATACTTATTAGCCTAGGATTAGGGACAACCCTAACTTTCGCCAGCTCCAACTGACITCTCGCATGAATGGGATTAGAAATTAACACT4101 CTAGCCATCATTOCACTTATAGCCAACAATCATCACCCACGAGCAGTAGAAGCTGCAACAAAATACTTCATCACTCAAGCCGCAGCCGCAGCCCTTTTAC4201 TATTTTCTAGCCTAATTAACGCCTGACAATCOGGSCABTGAATAATCCAAGACATATCAATACCCATATCTGCACTCATAAOCATTGCAATTGCCATTAA4301 ACTAGGCGTGGCOCCAGTACATTTCTGATTGOCTGAAGTTATACAAGGAATTAAATTAAATACAGGATTAATCTTAGCCACTTGACAAAAACTCGCACCT4401 CTGGCACTGCTHTATCAAATCTCTAACAACCTAATACCAGAACTCATAATMGCTHTAGGGTTAATATCGACAATCGTGGGGGATGAGGAGGACTTAACC4501 AAACACAAATCCGAAAAATCATAGCCTACTCATCTATTGCCCATCTAGGCTGAATTATCTCAATCATACACTTCATACCATCCCTCGCCATTATTAACCT
4601 AATTATATATATTATTTATGACAACAACAATATTCATAATTTTCAACACCCTAAACTCTACCACAATTAATGCCCTGGCCATTAATTGATCCAAATTCOCA4701 GCTCTCTCAGCCATCACCATACTCGCACTCCTATCATTAGGCGGACTCOCACCACTTTCAGGATTCCTGCCAAAATGACTTATTCTPCAAGAACTAACTA4801 ATCAAAATCTCGCACTAACCGCCACAGTAATAGCCCTATCAGCCCTACTTAGCTTATACTTCTATCTCCGACTCTCATATTCACTCACCACAACAATTAT4901 AСССААСАСАТАССААСАТАТАСТТААТТGAAACATCAAAACCAAAАТСАССТТТАТТСТАССААСААТААТААТТАТААСТАТСGССАТАСТСССААТС-trNA-Ala
5101 AGGATTTGGAGGACTHATCCCACATCTTATGAATGCAACTCAAACACTTTAATTAAGCTAAAACCCTACTAGATGAGTAGGCCTCGATCCTACATACGT $\leftarrow$ tRNA-Asn
5201 TTAGTTAACAGCTAAAAGCTUTAACCGGCAAGCTUCCACCTAGGCTTTCCTCCCGCCGAAACTCTAAAAGGCGGGAGGAAOSCCCGOCAGGCGTMAACCT $4-\mathrm{tRNA}$-Cys
5301 ACGRCTCAGGATTTGCAATCGTGCATGCAACTACACTACAAGGCTTGATAAGAGAAGGGATTAAACCTICCIUTACAGGGCTACAACCTGCCACTTAAAC $\mathrm{COI} \rightarrow$
5401 ACTCAGICATCTPACCTGTGACCATCACCCGCTGACTITTCTCAACAAACCACAAAGACATTGGCACCCTTGATTTAATCTTTGGIGOCTGAGCOGGAAT5501 AGTAGGAAOCGCACTAAGCCTCCTAATTCGCGCAGAACTAGGCCAACCAGGAGCOCTAATAGGAGATGACCAAATTTATAATGTTGTCGGCACTGCACAT5601 GCATTTGTAATAATTTTCTTTATAGTAATGCCAATCATAATCGGAGGATTCGGTAACTGACTTGTACCACTAATAATTGGAGCCCCAGATATGGCCTTCC

5701 CACGAATAAATAACATAAGTTTCTGATTACTYCCACCATCGCTTCTCCTGCTACTAACCTCTTCTGCAGTAGAGGCCGGTGTCGGAACAGGATGAACTGT

5901 ATTAACTTCATTACTACAATTATTAATATGAAACCGCCATCCACCTCACAATACCAAACCCOCCTATICGTATGATCAGTATTAGICACTGCAGTCCTCT

6001 TACTCCTATCCCTTCCAGTATTAGCCGCCGGAATTACAATACTACTAACAGATCGAAATTMAAACACTACCTTGTTTGACCCTGCTGGCGGAGGAGATCC

6101 AATTCTTTACCAACATTTATTCTGATTTTTTGGCCATCCAGAAGTATACATTTTMATTCTTCCAGGATTTGGTATAGTCTCTCATATCGTAGCTTACTAT

6201 TCAGGTAAAAATGAACCTTTCGGTTATATAGGAATAGTATGAGCAATAATAGCGATTGGACTCCTAGGATTTATCGTATGAGCCCACCACATATTTACAG

6301 TMGGTATAGACGTTGATACTCGAGCTTACTTCACTTCCGCCACAATAATTATTGCTATCCCTACTGGAGTTAAAGTATTTAGCTGACTCGCTACATTACA

6401 TGGAGGCGCTATTAAATGAGAAACCCCAATACTATGGGCTCTAGGCTTCATCTTCCTATTGACAGTAGGGGGACTAACCGGTATTATTTTAGCAAATTCA

6501 TCACTAGATATTATATTACACGACACATACTACGTGGTAGCACATTTCCATTATGITTTATCTATAGGAGCTGTCTTGGCCATTATGGGTGGITTGGTCC4000

3700

3800

3900
$\mathbf{W} \quad \mathbf{F} \quad \mathbf{P} \quad \mathbf{L} \quad \mathbf{F} \quad \mathbf{P}$
ACACTHCCTAGGACTAGCOGGAATACCCOGAOGCTACTCAGATTACCCAGATGCGTACACATTATGAAATHCTTTATCATCCATCGGGTCTATAATTTCC
6801CTAACOGCTGTCAITATATHTTTATTTATCCTCTGAGAGGCIHTOGCAGCCAAACGAGAAGTACAAACAGTAGAATTAACCCACACTAATGITGAATGAC$\begin{array}{llllllllllllllllllllllll}\mathbf{H} & \mathbf{G} & \mathbf{C} & \mathbf{P} & \mathbf{P} & \mathbf{P} & \mathbf{Y} & \mathbf{H} & \mathbf{T} & \mathbf{Y} & \mathbf{E} & \mathbf{E} & \mathbf{P} & \mathbf{A} & \mathbf{F} & \mathbf{V} & \mathbf{Q} & \mathbf{S} & \mathbf{P} & \mathbf{Q} & \mathbf{A} & \mathbf{R} & \mathbf{E} & *\end{array}$TTCACGОСIGCCCTOCCOCTTACCACAOGTACGAAGAACCAGCATTCGTCCAATCCCCACAAOCTCGAGAAAGGAAGGAAMMGAACCCCCTMTAAARCGOT
$\leftarrow$ trNA-Ser (UCN) tRNA-Asp $\rightarrow$7001 TTCAAGCCGACTGCATAACCATTCHGCCACTNTCTTATGAGATATTAGTLAAACCATAACATTGCCTTGTCAAGGCAAAATTACAGGMAAACCCCTGRA$\operatorname{COII} \rightarrow$7101 TATCITIATCTATGGCCCATCCCACACAACTAGGACTACAAGAOGCATCCTCTCCTATTATAGAAGACTACTACTCTTCCAOGATCACGCATTAATAACTGTATTMCTAATTAGCACACITGTACTTTATATTATTATAACCGCAGTTTCAACAAAACTAACCAACAAACATTTACTTGACGCCCAAGAAATTGAGATTG7301 TCTGAACAGITATACCAGCTCTAGTATTAATTGCTATTGCCCTACCGTCACTTCGAATOCTTTACCTAATAGATGAAATTAACGACCCTCATCTAACTAT7401 TAAAGCCACAGGACATCAATGATACTGAAGCTATGAATATACAGATTAOGACACATTAAACTTTGACTCATACATAATTCCCACACAAGATTTATTTACCA7501 GGACAATTCCGCCTCCITGACACAGACAATCGAATAGTTGTCCCCACCGGATCTCCTGTGCGCATACTCATTACAGCOGAAGACGTACTTCACTCATGAG7601 CAGTACCATCATTAGGCCTAAAAATAGATGCGGTGCCAGGACGACTTAATCAAACCACATTCATTGCCACTCGACCAGGAGTTTTCTTCGGTCAATGCTC7701 AGAAATTTGTGGAGCAAACCACAGCTHTATGCCAATTACAATCGAATCAGCGCCTGTTAAATATTPTTGAATCTTGATCITCATCAATACTAGCAGAATCAtRNA-Lys $\rightarrow$ATPase $8 \rightarrow$$\begin{array}{lllllllll}\mathbf{M} & \mathbf{P} & \mathbf{Q} & \mathbf{L} & \mathbf{N} & \mathbf{P} & \mathbf{N} & \mathbf{P} & \mathbf{W}\end{array}$7801 TCATLAAGAAGCTAATAGGGCATAGCATTLGCCTMUTAAGCTAAAAATAGGTGACTCCCAACCACCCTTAATGAATGOCACAACITAATCCAAACOCCTG$\begin{array}{llllllllllllllllllllllllllllllllll}\mathbf{F} & \mathbf{T} & \mathbf{I} & \mathbf{L} & \mathbf{I} & \mathbf{F} & \mathbf{T} & \mathbf{W} & \mathbf{A} & \mathbf{V} & \mathbf{F} & \mathbf{L} & \mathbf{T} & \mathbf{I} & \mathbf{L} & \mathbf{P} & \mathbf{N} & \mathbf{K} & \mathbf{V} & \mathbf{T} & \mathbf{S} & \mathbf{H} & \mathbf{K} & \mathbf{M} & \mathbf{P} & \mathbf{N} & \mathbf{E} & \mathbf{L} & \mathbf{L} & \mathbf{T} & \mathbf{K} & \mathrm{D} & \mathbf{P}\end{array}$7901 ATTTAOCATTTTAATTHTTACCTGAGCTGIATTTCTGACCATTTTTACCCAACAAAGTCACCTCACATAAAATACCTAATGABTTATTAACCAAAGACCCTATpase $6 \rightarrow$$\begin{array}{lllllllllllllllllllllll}M & T & L & S & F & L & D & Q & F & A & S & Q & \mathbf{S} & F & L & G & I & P & L & I & A & I & A\end{array}$
$\begin{array}{llllllllllllll}\mathbf{S} & \mathbf{N} & \mathrm{L} & \mathrm{L} & \mathbf{T} & \mathbf{E} & \mathrm{I} & \mathbf{W} & \mathbf{Y} & \mathbf{W} & \mathbf{P} & \mathbf{W} & \mathbf{H} & *\end{array}$
8001 TCAAAOCTCCTTACAGAAATCTGATACTGACCATGACATTAAGCTHTTTGGATCAGTTOGCAAGCCAATCCFTTTTAGGAATTCCACTAATCGCTATCOC8101 CATCCTAATTCCATGATATTATYTCCATCCOCCTACAAACGATGAATAAGCAACCGACTAATTACTHTTCAATCCTGATTTATMGCTCGCACCACTAAT8201 CAACTTATATTACCACTAAACACTGGTGCACATAAATGAGCTATAATTMTTAACTGCCCTCCTACTATTTCTAATAACTPTAAACCTCCTGGGTCTATTAC8301 CATATACATTTACACCAAOCACCCAACTATCAATAAATATAGCCCTGGCCGTACCACTATGACTAGCTACCGTATTAATTGGAATACGAAACCAACCAAC8401 ACACTCACTAGCCCATCTTCTTCCAGAAGGCACACCGACCCCACTAATCCCCATCCTAATTATTATCGAAACAATTAGCCTCTTTATTCGACCACTCGCT8501 CTGGGAGTACGACTAACAGCAAACTTAACCGCAGGCCACTTATTGATTCAACTAATCTCTACAGCAAOCTTTGTTATATTATCTAITATGCCAACAATCG8601 CCACACTCACATTTATTGTACTAGCTTTACTAACTATTCTAGAAATTGCAGTCGCAATGATTCAAGCTTACGTACTTGTCCTTMTATTAAGCCTGTATCTQ E N V COIII $\rightarrow$

8701 ACAAGAAAATGTCTAATGGCCCATCAAGCACACGCATATCATATGGTTGACCCAAGCCCATGACOCTTTAACAGGGGCAGTTGCCGCTTTACTGCTAACCT

8801 CCGGACTAGCAGTATGATTCCATTTCAAATCATTAACCCTACTAGCAATAGGTCTACTATTAATAATTTTTAACCATAATTCAATGATGACGAGACATCAT

8901 TCGIGAGGGAACATTTCAAGGACATCATACCCCACCTGTACAAAAAGGACTCCGCTATGGAATAATCTTATTCATTACATCCGAAGTATTCTTTTTCCTTT

9001 GGGTTTHTCTGAGCCTTTTACCATTCAAGTCTAGCCCCTACACCAGAATTAGGTGGCATTTGACCACCAACCGGTATTACACCTCTAGACCCATTCGAAG

9101 TTOCTCTTCITAATACAGCAGTACTACTAGCCTCTGGCGTAACTGTCACATGAACTCACCACAGCCTAATAGAAGGAAAACGAACTGAAGOCACACAAGC

9201 ACTAACCTTAACCATTTTATTAGGCTTATACTTCACCGCCCTTCAAGCAATAGAATATTATGAAGCCCCATTTACTATTGCAGACGGCGTTTACGGAACA

9301 ACTHTCITMGTAGCCACAGGCTTTCATGGACTTCATGTCATCATTGGCTCAACTTTTCTAGCAGGATGCTTATTTACGACAAATTCTTTATCACTTTAOCT

9401 CTTCCCACCACTITGGCTHTGAAGCTGCTGCATGATATTGACATTTCGTAGACGTAGTCTGACHTHTCCTHTATGIATCAATCTATTGATGAGGATCCTAtRNA-Gly $\rightarrow$$\begin{array}{llllllllll}M & \mathrm{~N} & \mathrm{~L} & \mathrm{I} & \mathrm{L} & \mathrm{M} & \mathrm{M} & \mathrm{I} & \mathrm{L} & \mathrm{I}\end{array}$
9501 ATTTWHCTAGTATTAAAAAGTACAAATGACTTCCAATCATTYAGTCTTGGCTAAATNCCAAGGAAAGATAATGAATHTAATCTTAATAATAATTTTAATII9601 TCCTCOCTAATCTCAACTATCCTAGCTATCGTCGCATTCTGACTOCCOCAAATAAATCCTGATATAGAAAAACTATCACCATACGAATGTGOCTTTGACC9701 CGCTCGGATCTGOCCGCCITCCATTCTCCATACGATTCTHCCTAGICGOCATTCTATTOCTCCTATTTGACCTAGAAATTGOCCTCCTHCITCCACTACC
$\mathbf{W} \quad \mathbf{S} \quad \mathbf{T} \quad \mathbf{H} \quad \mathbf{I} \quad \mathbf{D} \quad \mathbf{P}$9801 ATGAAGCAOCCACTTAGACCCCACTCTTATACTAATATGAGCATTTACCATTPATTATTCTCCTGACAATTGOCCTAATTTACGAATGACTTCAAGGTGGA
L E W A E$\mathrm{NADH} 4 \mathrm{I} \rightarrow$
tRNA-Arg $\rightarrow$ ..... M T H I M
9901 THAGAATGAGCAGAATAAGLCTCTAGICCAACGNAAGATVYTTAATYWCGGCTWAATAGATXGTGGCTCAAATCCACAGAGACCWAATGAOCCATATCAI10001 ATICACCTYCTCAACCGCCITWATACRAGGATTATCAGGCCTAACTYTCAATCGCACTCACCTACFCTCTGCCCTGCTATGTHPAGAGGGTATAATATTA10101 TCCITATVTATTGCCCTAGCAATATGATGCACCCAAAATGAGACTATAATATTTCTCTTOCGCCCOCCTACTATTACTAGCCCTCTCGOCGTGCGAAGCAG
NADH $4 \rightarrow$
$\begin{array}{llllllll}\mathbf{M} & \mathbf{L} & K & \mathbf{L} & \mathbf{I} & \mathbf{I} & \mathbf{P} & \mathbf{T}\end{array}$10201 GCCTCGGCCTAAGCCTACTTGTCGCCACTGCOCGCGCCCACGGGTCTGATCACCTHCAAAACCTTAACCTCCTACAATGTTAAAACTACTAATPOCAACCATTATACTATTCCCAATAATTTGGACACTAAATCCTAAATGATTGTGATCAGCTACCACAACACATAGCTTAATTATTGCTTCCTTATCTCTCACACTAT10401 TTAAATGCTACTCCACAACCCAATGATCAAATCTTAATTATATACTAGCCAOCGACATAATCTCCACOCCTTTAATTATCCTAACCTGCTGACTACTTCC10501 ACTAATAATTATTGCTAGOCAAAACCATATATCCACGGAACCCATCAACCGACAACGAAGTTACATTACACTACTAGTATCCCTGCAAGCCCTACTTATT10601 ATAGCCITCAGCGCCACAGAAATTATTTTHATTTTATATTATATTTGAATCCACCTTAATCCCTACCCITATTATTATTACACGCTGAGGAAACCAAACAG10701 AAOGTCTTAACGCTGGCATCTACTTCCTATTTTATACTCTAGCCGGCTCACITCCATTACTAGTTGCOCTCCTATATTTATATAACACAGCCGGCTCGCT10801 GTCATTHATTTCAATAAACTPAATCTCTATCOCCCCTAACACTTTGAACCAACACTTTCTTATGAGTAGCCTGCGTAACCGCCTTCCTAGTAAAAATACCT10901 TTATACGGAGTTCATCICTGACTCCCCAAAGCCCATGTAGAAGCOCCAGTTGCCGGATCAATAATTCTTGCOGCCATCCTACTAAAATTGGGCGGATACG11001 GAATAATTCGAATAAOCATTATACTAGAACCTGCAACCAAATCATTAGOCTACCCATTGCATTATTCTTGCATTATGAGGAATCATTATGACAGGGTCAATTTGTATACGACAATCAGACATAAAATCCCTAATCGCCTACTCCTCAGTCAGCCATATAGGACTAGTTGCCTOCGGCATCCTAATCCAAACCACATGAGGT11201 TTTACAGGAGCAATTATCTTAATAATTGCCCACGGGTTAACATCATCAOCCCTATTCTGCCTAGOCAACACOGCCTACGAACGAACACACTCCCGAACTCTCCTCCTAGCACGAGGAATACAAATCATCTTACCATTAATAGCGACTTGATGATTTATTATAAGTCTAGCAAACATAGCTTTACCTCCACTTCCCAACCT11401 AATAGGAGAACTAATAATCCTAGTATCTATATTTAACTGATCCAACTGAACTATTTTTATTAACAGGTACAGGAACCCTAATTACAGCTAGTTACTCACTA11501 TATCTHTATATATCATCTCAACGAGGACCAACCCCCAACAACCTAACCTTTATAGAACTTTOCCACACCCGAGAACACCTCCTACTAACCTTACATATTA11601 TTCCAATTATCCTCCTAATAATTAAACCTGAACTTATCTGAGGATGATGTTGATGTMAATATAGTCTAAATAAGACATTAGATTGIGATMCTAATAATAG
tRNA-Ser (AGY) $\rightarrow$ tRNA-Leu11701 AAGTTAAACTCTTCTTATTGACCGAGAGATGCTCGCGCACCAAGAACTGCTAATTCTTATTACCTTGGHTAAACTCCAAGGATCACTCGATGCTHMCAAA
NADH-5 $\rightarrow$(CUN) $\rightarrow$$M \quad S \quad I \quad S \quad Q \quad S \quad Q \quad M \quad F \quad M \quad T \quad C$11801 GGATAATAGATATCCATTGGTCTTAGGAACCAAAAACTCTMGGTGCAAATCCAAGTGAAAGCTATGTCTATCTCCCAATTATCACAAATATTTATAACAT11900GССTCTCCCTAACAATAATTATTCTCATCCTACCAATCACGTTTTCCTTCATTACGAAACCATCAAACAAATGACCTTTCCAAGTCAAAAATGCTGTAAAATTATCCTTCTTTGTAAGCCTAATTCCATCAATTACTTTGCTTAAACTTAAATCTCCAATCCTTTACCATCTACTACCAATGATTTTCTATTTCATCCACATACACACTGACATTTTAATTAACCGATTCTTTTAAATACCTATTAACATTTCTAATTGCCATAATAATTCTGGICACAGCTAATAACATATTCCAACTTTTT12301 TATCGGCTGAGAAGGAGTAGGAATCATATCATTCCTACTAATTGGGTGATGATATGGCCGCGCCGATGCCAACATAGCCGCATTACAAGCTGTAATTTAT1200012400
NADH 3 $\rightarrow$
 12401 AATCGTGTAGGAGACATCGGTCTCATAATAACAATATCTTGACTTTTTAATCAACACTAACTCATGAGATATTCAACAACTATTTGGCCTTACAAAAAATA
 12501 TAGACACAACACTHCCTGCABOCGGACTGCTACTAGCAGCAACAGGCAAATCCGOCCAATYTGGCCTMCACCCATGACTACOCGCAGCAATAGAAGGTCC T $\mathbf{P}$
12601 AАСТССАGTCTССОСССТАСTTCACTCCAGCACAATAGTTGTTGCCGGAATCTTOCTGTTAATCOGCCITCACCCACTCATTGAAAACAATAACAATATC
 12701 CTTACCGCTGCACTCTGCCTAGGCGCAATTACTACOCTCTTCACCGCCACCTGCGCOCTGACACAGAATGACATCAAAAAAATCGTTGGATHCTCTACAT

12801 CCAGTCAACTAGOCCTAATAATAGTAGCAATTGGACTTAACCAACCCCAATTAGCATTTCTTCACATCTGCACCCACGCATTTTHCAAAGCCATACTCTT
 CCTATGTTCTGGATCAATTATCCACTCCTTGAATGACGAACAAGACATCCGTAAAATAGGCGGAATTAATAAAACACTCCCATTAACCTCCTCCATGCTTA
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 13101 GAGCCCTAGTACTAACACTAATCGCCACCTCATTCACCOCCGTGTACAOCTTACGCATCATCTATTTTGTCTTAATAAATCATCCTCGIACCOTOCCOCT
 13201 CTCTCOCGTAAATGAAAATAATCCCTTAATTGCTAATCCAATTAAACGCCTTGCATGAGGAAGCATTATTGCCGGATTAATTCTITTGTCAATATATTCTT
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13501 ATTACCGCTAATACTAGGACAAATCTCTGCTACACAAATATCAGACCAACTATGAATAGAGAAATTAGGACCAAAAGGCATTGCCCACACACAACTTCTC

13601 GTTACOCAAAAAATTACACATGTACATAAAGOCTAATTAAAACATACTTATCAATMATAATACTATCAATCATMATTATCACAATTATCATTATAATTA
 13701 CCTAACCGCACGTAAACAACCACGAGAATATCCTCGAGTTAATTCTAGCACAACTAATAAAGCTATTAATAATACCCACCCAGCAATAAACAACATATAA
 13801 ССТССАGСАITATATACCTCTGСТАССССААСАТАТСGССGСАТGGССССААААТАТТОСТСААСАТСАТСАААТААТТСТАТАТССССААСАААТGСТА
 13901 CССАТGСTACAATAACTAAAAACATATATACTAATACATAAGAAAAAACCTCCCAAGAACCOCACGCATTAGGATAAGGCTCCGCTGCCAATGCAGGAGA

14001 ATAAGCAAATACTACTAACATCCCTCCCAAATAAATTAAAAAAAGAATTATAGATAAAAAGTTATACCACATTGCATTAACATTCCACAOCCAACTCCC

$$
\leftarrow \mathrm{NADH} 6
$$


14101 GCTCCAACTATCAACCCAAAAGCTGCAAAATAAGGAGAAGGATTAGATGCTACTGCAATTAAACTAATTAAAAATATTICAGAAAAACTACCATAAGTA Cyt b $\rightarrow$

14201 CCATAATNCTMGCCAGGACTYULACCAGGACTAATGACTTGAAAAATCACCGTTGTAACTCAACTACABAAATCAAATGGCAATCATCCOCAAAAOCCAC
 14301 CCTCTAGCAAAAATTATTAACAGTGCATTCATTGACCTACCAGCOCCATCAAACATCTCATCATGATGAAACATAGGCTCCCTACTTGGATTATGCCTAA

14401 TCGCACAGATTATCACAGGACTATTCTTAGCCATACATPACGINTCCGATATTAACTCAGCTHTCTCCTCAGTCOCACACATCTGCCGTGATGTAAACTA 14500

14501 TGGATGACTAATCCGAAACTHCCACGCAAACGGAGCATCCCTATTCTTTCATTTGCATMTACTTACATATCGOGCGCGGTCTATATTATGGATCCTACCTC
 14601 TACACAGAAACCTGAAACATAGGGGTAATTCTACTCCTCCTCACCATAATAACCGCATTTGTAGGTTATGTCCTTCCATGAGGGCAAATATCCTTCTGAG

14701 GTGCCACAGTTATTACCAATCTTCTATCAGCAATCOCATACATCGGAGATAOCTTAGTACAATGAATCTGAGGAGGGITMTCAGTTGATAAACCAACACT

14801 TAOCCGATTCTTTGCATTOCACTTCATTCTTOCCTTCGCAATCGCAGCAGCGTCCCTCGTCCATATTGTATTTCTTTCATGAAACAGGGTCAAACAACOCA

14901 GTAGGAATTAATTCCGATGCAGACCAAATTCCATTTCAOCCTTACTTTACCTTTAAAGACCTACTAGGCTTCATTATCTTACTACTAATTATTATTATAT

15001 TAGCATTACTTTCACOCAACCTATTGAATGAOCCAGGAAATTTTACTCCAGCCAACCCCTTAATTACCCCCCCTCATATTAAACCTGAATGATATTTOCT

15101 ATTTGCCTACGCAATTCTACGTTCTATTCCCAACAAACTAGGAGGAGTGTTAGCCCTACTATTCTCCATCCTTATCCTAATGTTAGTACCAITACTCCAC $\mathbf{T} \quad \mathbf{S} \quad \mathbf{K}$
15201 ACCTCAAAAATTCGCAGCGCCACATTCCGCCOGCTATTCAAAATTPACACTCTGAATTCTCGCAGCAGACGTCCTCATCTTAACATGAATTGGAGGACAAC

15301 CAGTAGAAGACCCGTACATTATTATTGGOCAAGCCGCCTCGATCCTTTACTTCTTAATTTTCTTAGTACTTATACCACTATCAGGTTGACTAGAAAACAA

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<td style="text-align: center; border-right-style: solid !important; border-right-width: 1px !important; border-bottom-style: solid !important; border-bottom-width: 1px !important; border-top: none !important; width: auto; vertical-align: middle; " class="_empty"></td>
</tr>
</tbody>
</table>
<table-markdown style="display: none">|  |  | 1550 |
| :---: | :---: | :---: |
|  | $\leftarrow$ tRNA-ProControl Region $\rightarrow$ TAS-1 |  |
| 15501 | THWAACTCCTGCCTNTLACTCCCAAAGCTAAAANCTLACTTAAACTACTCTMTGATCATGTATGTATATAAGACACAATATTTAAACAAACATTACAT TAS-2 | 1560 |
| 15601 | TATCCOCTACATAAAOCATATAACTATTAATCAGGTATTACACCCACCATATCAAAGTACACTACCCTGGTCACTACTACCOCACCTOCCOCCCCAAAAT $\qquad$ | 1570 |
| 15701 | TATATATCCCCAATAATTAATAACAA | 1580 |
| 15801 | ATATGCTCCATGAGCICCATGAACAACAATTPATCAAGAAATACCTTACTAAAACCACAACATGGATACTAACTACAGCCITATAAGTACATATATA | 900 |
| 15901 | TACTTAACACCTATTAAGCCAGGAATATACATGTAATYTACCATTAAATTATGTACAGCATTCAGGAGAGTCAITCGTTTTTCATATrTATCAACAATATM | 1600 |
| 16001 | CCATAATAACAGTCTCOICCGGATTGTAACCACCATCCTAAACGGTTCCACCCTACCCGACCATCATTATAATCTAAGACTTATTAMCCGTGATACACT CB—D | 1610 |
| 16101 | AACGITCCATTCCAGAACATTHGGTTCCTAATTCAGGCACATAAATAGATAATTOCCTATACOTGATATTATCGAGGCTATAGTGACTGTACGGAACATG | 1620 |
| 16201 | GGTCTCCTGGCACECGACATGGTTPAACAGMCCTCGTACGTICOGCACTGOGGTTGTTHAACCTGATCTGCCTCATTAGCCHTMTICOTCOTTCACITT | 16 |
|  | CSB-I |  |
| 16301 | GCC |  |</table-markdown></div> <br> 16401 TTYTAOCATTAATTATTCTCTCAAGAGCATAACTACTTTTACATCCCTHGGTGCTTTCTTAATAAATTAAAATTTTAAAACGCGAAAATAGCCTAAAACC 16500 CSB-II CSB-III <br> 16501 CCCTACCCOCTTAAAAAAGACGTTTTTGGACTITTTGGCCCOGTAAAACCCOGAAAACAGGAAAGIGCCTAGAAACGCAAGAGAATTGTGTTGCTGCAAA 16600 <br> 16601 TCATCITGCAGIGITGCACTACGA 16624 

Figure 3.- Continued
than to those of either the lungfish or the lamprey (Table 4).

All protein encoding genes in the bichir mitochondrial genome use the initiation codon ATG except for COI, which uses GTG. The use of GTG for initiation of translation in COI is shared with other fish mitochondrial genomes that have been sequenced completely and with the chicken mitochondrial genome. Three genes (ATPase 8, ND4L, and ND5) use TAA as stop codon, eight genes (ND1, ND2, COII, ATPase 6, COIII, $N D 3, N D 4$, and $c y t b$ ) use the incomplete stop codon T , and the remaining two genes ( $C O I$ and $N D 6$ ) use an AGR stop codon (Table 3). Of the complete vertebrate genomes sequenced, no ray-finned fish uses AGR as a stop codon, whereas lamprey, lungfish, frog, chicken and mammals do use AGR stop codons.

The overall codon usage of the 13 protein genes of the bichir mtDNA genome shows a strong bias against guanine at third positions. The anti-G bias in bichir is the second strongest ( $4.4 \%$ ) after lamprey ( $3.8 \%$ ), while other fish and frog are less strongly biased (Meyer 1993) (Table 5). As seen in other mitochondrial genomes (Naylor et al. 1995) pyrimidines are overrepresented compared with purines in the second codon position (percent $\mathrm{C}+\mathrm{T}=68.3$ ) due to the hydrophobic character of proteins encoded by mitochondrial genes.
Transfer RNA genes: The bichir mitochondrial genome contains 22 transfer RNA genes that are interspersed between ribosomal RNA and protein coding genes. The transfer RNA genes range in size from 67 to 75 nucleotides. All bichir tRNA genes can be folded into conventional cloverleaf secondary structures. As seen in other vertebrate tRNAs formation of G-U, and other atypical pairings are found in the stem regions. All proposed cloverleaf structures, except that of $\mathrm{tRNA}^{\operatorname{Ser}(A G Y)}$ contain 7 bp in the amino acid stem, 5 bp
in the TYC stem, 5 bp in the anticodon stem and 4 bp in the DHU stem. In most vertebrates $\mathrm{tRNA}^{\text {Ser(AGY) }}$ has a reduced DHU arm. The bichir tRNA ${ }^{\operatorname{Ser}(A G Y)}$, similar to that of the lamprey $\mathrm{tRNA}{ }^{\text {Ser(AGY)}}$, has no recognizable DHU stem and loop. Rainbow trout, carp and cod on the other hand posses a $\operatorname{tRNA}{ }^{\text {Ser(AGY) }}$ which is much less truncated (ZARDOYA et al. 1995).

Ribosomal RNA genes: The 12 S and 16 S ribosomal RNA genes in bichir are 950 and 1655 nucleotides long, respectively. They are situated as in other vertebrates between $t R N A^{P h e}$ and $t R N A^{L e x(U U R)}$ and are separated by the $t R N A^{V a t}$. The ribosomal RNA genes follow the same trend as the protein encoding genes and show greater sequence similarity to ray-finned fish than to lamprey or lungfish (Table 4).

Noncoding regions: The control region, which is the largest noncoding region found in the bichir mtDNA, is 1068 nucleotides long and is situated between the $t R N A^{\text {Pro }}$ and $t R N A^{\text {Phe }}$ genes. The control region of vertebrates contains the site of initiation for heavy strand replication as well as promoters for the transcription of both the light strand and the heavy strands. The bichir control region has an overall base composition that is rich in $A$ and $T$ (percent $A+T=64.3$ ). Between positions 16,100 and 16,300 (Figure 3), a G-rich region is found within which the central conserved block $D$ (SOUTHERN et al. 1988) was identified. This sequence block is conserved between bony fish and mammals (Lee et al. 1995), but has not been found in the lamprey control region (Lee and Kocher 1995). The conserved sequence blocks CSB-II and CSB-III, which are thought to be involved in the generation of the RNA primer needed for initiation of heavy strand replication (WAL berg and Clayton 1981), are present in the right domain of the bichir control region. Similar to lungfish (Zardoya and Meyer 1996a) and frog (Roe et al. 1985),

TABLE 3
Localization of features in the mitochondrial genome of Polypterus ornatipinnis

| Feature | From | To | Size, bp | Codon |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
|  |  |  |  | Start | Stop |
| tRNA-Phe | 1 | 71 | 71 |  |  |
| 12 SrRNA | 72 | 1021 | 950 |  |  |
| tRNA-Val | 1022 | 1092 | 71 |  |  |
| 16 Sr rNA | 1093 | 2747 | 1655 |  |  |
| $t R N A-l e u(U U R)$ | 2748 | 2822 | 75 |  |  |
| NADH 1 | 2823 | 3780 | 958 | ATG | T- |
| tRNA-Ile | 3781 | 3853 | 73 |  |  |
| tRNA-Gln | 3854 | 3920 | 67 (L) |  |  |
| tRNA-Met | 3921 | 3989 | 69 |  |  |
| NADH 2 | 3990 | 5025 | 1036 | ATG | T- |
| tRNA-Trp | 5026 | 5094 | 69 |  |  |
| tRNA-Ala | 5100 | 5168 | 69 (L) |  |  |
| tRNA-Asn | 5170 | 5242 | 73 (L) |  |  |
| $t \mathrm{RNA}$-Cys | 5278 | 5346 | 69 (L) |  |  |
| tRNA-Tyr | 5347 | 5416 | 70 (L) |  |  |
| COI | 5418 | 6974 | 1557 | GTG | AGG |
| $t$ RNA-Ser (UCN) | 6966 | 7036 | 71 (L) |  |  |
| $t$ RNA-Asp | 7039 | 7107 | 69 |  |  |
| CO II | 7111 | 7801 | 688 | ATG | T- |
| tRNA-Lys | 7802 | 7874 | 73 |  |  |
| ATPase 8 | 7875 | 8042 | 168 | ATG | TAA |
| ATPase 6 | 8033 | 8714 | 682 | ATG | T- |
| CO III | 8716 | 9499 | 784 | ATG | T- |
| tRNA-Gly | 9500 | 9570 | 71 |  |  |
| NADH 3 | 9571 | 9916 | 346 | ATG | T- |
| tRNA-Arg | 9917 | 9986 | 70 |  |  |
| NADH $4 L$ | 9987 | 10283 | 297 | ATG | TAA |
| NADH 4 | 10277 | 11654 | 1378 | ATG | T- |
| tRNA-His | 11655 | 11723 | 69 |  |  |
| tRNA-Ser (AGY) | 11724 | 11789 | 66 |  |  |
| tRNA-Leu (CUN) | 11792 | 11863 | 72 |  |  |
| NADH 5 | 11864 | 13705 | 1842 | ATG | TAA |
| NADH 6 | 13701 | 14204 | 504 (L) | ATG | AGG |
| tRNA-Glu | 14205 | 14273 | 69 (L) |  |  |
| CYT b | 14277 | 15417 | 1141 | ATG | T- |
| tRNA-Thr | 15418 | 15488 | 71 |  |  |
| tRNA-Pro | 15489 | 15556 | 68 (L) |  |  |
| Control region | 15557 | 16624 | 1068 |  |  |

a putative CSB-I with a reduced motif of GACAT can be identified at position 16,398 (Figure 3).

At the $3^{\prime}$ end of the control region, the palindromic sequence motives TACAT and ATGTA are repeated four and three times, respectively. These motives have
also been found at the $3^{\prime}$ end of other vertebrate control regions and are thought to act as a recognition site for the termination of heavy strand elongation by forming stable hairpin-loop structures (SACCONE et al. 1991). In the left domain, we could also identify four putative termination associated sequences (TASs) based on the consensus mouse TAS sequence proposed by Doda et al. (1981) (Figure 3).

The origin of light strand replication $\left(\mathrm{O}_{\mathrm{L}}\right)$ is located between the $t R N A^{A s n}$ and $t R N A^{C s s}$ genes and is 46 nucleotides in length (Figure 3). The sequence has the potential to form a stable stem-loop structure with 11 bp in the stem and 12 nucleotides in the loop. The conserved motif $5^{\prime}$-GCCGG- $3^{\prime}$ at the base of the stem within the $t R N A^{C y s}$ gene, which has been shown to be involved in the transition from RNA synthesis to DNA synthesis (Hixson et al. 1986), is also conserved in the bichir. The most interesting feature of the bichir $\mathrm{O}_{\mathrm{L}}$ is a stretch of T-rich sequence in the loop. This stretch of T-rich sequence in humans has been shown to be involved in the synthesis of a RNA primer for initiation of light strand replication (WONG and Clayron 1985). All mammalian mtDNAs sequenced so far have this T-rich sequence, whereas all other fish mtDNAs sequenced have a stretch of cytosines or, as is the case for lungfish, a stretch of cytosines and thymines (Zardoya and Meyer 1996a).

Phylogenetic analyses of bichir mitochondrial genome: To phylogenetically correctly place the bichir among vertebrates, and especially to assess its relationship to ray-finned and lobe-finned fish, we used the mitochondrial genes of the following species for phylogenetic analyses: human (Anderson et al. 1981), whale (Arnason and Gullberg 1993), marsupial (Janke et al. 1994), chicken (Desjardins and Morais 1990), frog (Roe et al. 1985), lungfish (Zardoya and Meyer 1996a), rainbow trout (ZARDOYA et al. 1995), carp (Chang et al. 1994), loach (Tzeng et al. 1992) and lamprey (Lee and Kocher 1995). Protein-encoding genes were aligned and gaps were introduced according to the deduced amino acid sequences. Variation among the 13 protein coding genes was mainly found in the carboxyl end of the polypeptides and in few cases in the amino end. However, the central core of the mitochondrial proteins was found to be highly conserved. Therefore, ambiguous alignments at $5^{\prime}$ and $3^{\prime}$ ends of protein-coding

TABLE 4
Percentage nucleotide similarity of bichir mitochondrial genes against their homologues of other available fish

|  | 12 S | 165 | NDI | ND2 | ND3 | ND4 | ND4L | ND5 | ND6 | ATP6 | ATP8 | COI | COII | COIII | CYT 6 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| bichir-lamprey | 68.3 | 66.8 | 66.8 | 57.7 | 68.4 | 61.9 | 57.4 | 61 | 56.9 | 63.9 | 50.6 | 77.6 | 71 | 74.4 | 65.6 |
| bichir-rayfinned fish ${ }^{2}$ | 73.9 | 71.2 | 71 | 63.7 | 70.5 | 69.4 | 67.7 | 67.3 | 61 | 70.9 | 61.7 | 78.5 | 75.3 | 77.1 | 72.2 |
| bichir-lungfish | 71.2 | 68.9 | 67.4 | 61.9 | 66.2 | 65 | 58.6 | 62.9 | 59.3 | 66.7 | 57.1 | 76.2 | 73.8 | 75.8 | 69.3 |

[^2]TABLE 5
Base composition for the 13 protein-coding genes of fish and an amphibian

|  |  |  |  |  |  |
| :--- | :---: | :---: | :---: | :---: | :---: |
|  | Codon position | A | G | C | T |
| Lamprey | 1 | 30.4 | 22.6 | 22.9 | 24.1 |
|  | 2 | 19 | 12.9 | 26.5 | 41.6 |
| Polypterus | 3 | 41.3 | 3.8 | 21.5 | 33.4 |
|  | 1 | 29.9 | 23 | 23.7 | 23.4 |
|  | 2 | 18.9 | 12.8 | 27 | 41.3 |
| Trout | 3 | 42.7 | 4.4 | 27.8 | 25.1 |
|  | 1 | 25.4 | 26.4 | 26.8 | 21.4 |
|  | 2 | 18.2 | 13.8 | 27.7 | 40.3 |
| Carp | 3 | 33.4 | 8.9 | 33.9 | 23.8 |
|  | 1 | 27.1 | 25.9 | 26.4 | 20.6 |
|  | 2 | 18.5 | 14 | 28.2 | 39.3 |
| Loach | 3 | 44.2 | 5.9 | 31.3 | 18.6 |
|  | 1 | 27.2 | 26.4 | 25.6 | 20.8 |
|  | 2 | 18.5 | 13.7 | 27.7 | 40.1 |
| Lungfish | 3 | 35.8 | 9.6 | 34.6 | 20 |
|  | 1 | 27.6 | 23.6 | 25.4 | 23.4 |
|  | 2 | 18.4 | 13.3 | 27.2 | 41.1 |
| Frog | 3 | 34.7 | 8.4 | 28.5 | 28.4 |
|  | 1 | 29.9 | 21 | 23.3 | 25.8 |
|  | 2 | 20.5 | 11.6 | 27.2 | 40.7 |
|  | 3 | 41.2 | 6.5 | 22.3 | 30 |

Values are expressed as percentage.
genes were excluded from the phylogenetic analyses. The analyses were performed using nucleotide sequences and third codon positions were excluded since they are largely saturated with back mutations at this level of phylogenetic inquiry (e.g., CAO et al. 1994; Russo et al. 1996; Zardoya and Meyer 1996a). Similarly, all tRNA genes were aligned taking their secondary structures into account. DHU and T $\Psi \mathrm{C}$ arms were omitted due to ambiguity in alignments, and all positions were weighted equally. Moreover, both rRNA genes were aligned also taking their secondary structures into account and questionably aligned sequences were omitted from the analyses. For rRNA genes, all positions were weighted with a transition/transversion ratio of 2:1 to account for the faster rate for transitions over transversions in these genes (e.g., Meyer 1993; Ortí and Meyer 1996). In all analyses, gaps in alignments were treated as missing data.
In their investigation of lungfish relationships among vertebrates, ZARDOYA and MEYER (1996a) found that if the lamprey is used as outgroup, vertebrate trees with unexpected topologies and low bootstrap values are found regardless of which phylogenetic method is used. The same results are obtained when bichir is added to the analysis and lamprey is used as outgroup. It seems that too many multiple substitutions might have accumulated along the fast evolving lamprey lineage, hindering the recovery of correct phylogenetic relationships among vertebrates (Russo et al. 1996; Zardoya and Mexer 1996a,c). We used the same taxa that were included in the Zardoya and meter (1996a) study to


Figure 4.-Fifty percent majority rule bootstrap (Felsenstein 1985) consensus tree of jawed vertebrates based on 1000 replications. Two data sets were subjected to MP (bootstrap values above branches) and NJ (bootstrap values below branches) analyses. The first data set comprises a combination of all mitochondrial genes (bootstrap values upper of each pair of numbers). The second data set includes all mitochondrial protein coding genes combined (bootstrap values lower of each pair of numbers). ML analyses with the same two data sets yielded the same topology. Bichir, rainbow trout, carp, and loach were used as outgroup for all analyses.
investigate the performance of the bichir as outgroup in resolving vertebrate phylogenetic relationships. The phylogenetic analyses were performed using a data set of all mitochondrial genes combined and individual data sets of combined protein coding genes, tRNAs or rRNAs only. All three commonly used phylogenetic methods (MP, NJ, ML) arrived at identical and strongly supported topologies when bichir, rainbow trout, carp and loach were used as outgroup taxa and the combined set of all mitochondrial genes was analyzed (Figure 4)

With the separate protein data set, the expected phylogeny was also obtained with all three phylogenetic methods used (Figure 4). Strikingly, the separate data sets for tRNAs or rRNAs were unable to resolve the expected vertebrate relationships. In the case of the rRNA data set, the MP and ML analyses placed frog and lungfish as the sistergroup of teleosts and this grouping was supported by a bootstrap value of $56 \%$ (MP). NJ analysis of this data set arrived at a tree that placed frog and lungfish into a monophyletic group basal to tetrapods, this clade was supported by a bootstrap value of $52 \%$. For the tRNA data set, all three phylogenetic analyses (MP, NJ, and ML) arrived at unexpected trees in which the lungfish was basal to teleosts and tetrapods. This topology was supported with bootstrap values of 74 and $57 \%$ for MP and NJ analyses, respectively. In the parsimony analyses of the tRNA and

TABLE 6
Statistical confidence of maximum likelihood trees

| Tree topology | tRNAs |  |  | rRNAs |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | $\log l$ | $\Delta l i$ | SE | $\log l$ | $\Delta l i$ | SE |
| 1. (bichir,(trout,(carp,loach)),(lungfish,(frog, (chicken,(marsupial, (whale,human) )) ) ) | -8746 | -6.7 | 6.3 | -11,004 | $-11.2$ | 16.6 |
| 2. (bichir, ((frog,lungfish), (trout, (carp,loach)) , (chicken, (marsupial, (whale,human))) | -8762 | -22.6 | 11 | -10,992 | - | - |
| 3. (bichir,lungfish, ((trout, (carp,loach)), (frog, (chicken, (marsupial, (whale,human)) )) ) | -8739 | - | - | -11,012 | -19.4 | 17.1 |

$\Delta l i$ represents the difference in log-likelihood between the best tree ( $i$ ) obtained for each data set and the expected tree (tree \#1). SE is the standard error. Trees are declared significantly different when the difference in log-likelihood is larger than 1.96 standard error.
rRNA data sets, only a few more steps are needed to recover the expected topology (Figure 4), suggesting that the shortest trees obtained for those two data sets are poorly supported and not statistically different from the expected tree. This finding from the MP analysis was confirmed with ML when standard errors of the difference in log-likelihood between the ML tree given by each data set and that of the correct tree were calculated by the formula of Krshino and Hasegawa (1989). Both the rRNA data set and the tRNA data set exhibited log-likelihood ratios for the expected tree that were not significantly lower than those of the best tree obtained in each case (Table 6). This suggests that the expected tree (Figure 4) cannot be statistically ruled out for the tRNA and rRNA separate data sets. This conclusion is also confirmed by the higher bootstrap values (with MP and NJ ) obtained for the combined data set of all mitochondrial genes compared with the bootstrap values (MP and NJ ) obtained for the separate data set of all protein-coding genes (Figure 4). This stronger support of the expected topology is due to the addition of the phylogenetic information contained in the rRNA and tRNA data sets to that contained in the proteincoding data set.
Phylogenetic position of bichirs: Analyses of the combined protein-coding genes by MP, NJ, and ML
(Figure 4) rule out the hypothesis that the bichir is a sarcopterygian (Figure 1a). Without the addition of an appropriate, and yet unavailable, outgroup, it remains, however, difficult to distinguish between the two hypotheses of bichir as a third major bony fish lineage (Figure 1b) or the birchir as a basal actinopterygian (Figure 1c). One problem in assessing polypterine relationships is that the bichir is one of the most basal vertebrate taxa for which the complete mitochondrial genome is currently available. Lamprey has been shown to be unable to resolve vertebrate relationships and is too distant to be used as outgroup (Russo et al. 1996; Zardoya and Meyer 1996a, c). Therefore, the phylogenetic position of the bichir can only be determined in the future if members of the Chondrichthyes (sharks, rays and chimaeras) are used as outgroup and a sturgeon mitochondrial genome has been sequenced.

The Cytochrome b gene has been sequenced for both the white shark (Carcharodon carcharias) and the tiger shark (Galeocerdo cuvier) (Martin and Palumbi 1993). Additionally, the sturgeon cyt $b$ sequence is also available (Brown et al. 1989). To further test polypterine relationships, the same analyses as described above were performed with this gene using the same 10 taxa as above plus sturgeon and the two sharks. This data set was analyzed with MP, NJ and ML using both nucleotide sequences (third positions


Figure 5.-Majority rule bootstrap (FelsenSTEIN 1985) consensus trees of the cytochrome b data set. (A) Tree was obtained from MP analysis. (B) Tree was obtained from NJ analysis. Numbers above branches indicate bootstrap values ( 1000 replicates). ML analysis arrived at the same topology shown for tree A .


Figure 6.-Phylogenetic hypothesis and pectoral limb evolution. The basal (b) and radial ( r ) bones are indicated in each of the fin/limb schematics. Only up to three basals and radials are indicated, often their number is larger. The homology of some basal bones, e.g., in the birchir and the bowfin (Amia calva) is disputed. The homology, and numbers of some of the axial elements between ray-finned and lobe-finned fish is ambiguous (from Romer and Parsons 1977).
were excluded) and amino acid sequences and the sharks were declared outgroups. MP and ML analyses place the bichir as sistergroup to all other ray-finned fish included in the analysis; i.e., the bichir is placed basal to sturgeon, supporting the view that bichir should not be placed together with the Acipenseriformes (Table 1) into the Chondrostei (contra Nelson 1994). However, the bootstrap value for the node combining bichir with the other ray-finned fish in MP analysis is low ( $36 \%$ ), leading to an unresolved trichotomy between bichir, ray-finned fish and lobe-finned fish in a $50 \%$ majority rule consensus tree (Figure 5a).
Interestingly, the N analysis places the bichir as the sistergroup to lungfish and tetrapods supporting the
theory that the bichir might be a distinct piscine lineage (Brachiopterygii, sensu Bjerring 1985) that is more closely related to sarcopterygians than to actinopterygians (Figure 5b). This tentative topology is only weakly supported by a low bootstrap value of $62 \%$.

For all three phylogenetic methods (MP, NJ and ML), the relationships between lungfish and frog are not convincingly resolved with the cytochrome b data set. The discordant results and the low bootstrap values for generally well-established tetrapod relationships support the notion that single genes often are unable to obtain the correct phylogeny among such lineages that diverged $>400$ mya (CaO et al. 1994; Russo et al. 1996; Zardoya and Meyer 1996a,c).

Our results show that the bichir is a reliable outgroup to assess relationships among jawed vertebrates, leading to well-supported topologies when large data sets (all mitochondrial genes combined or at least all protein coding genes combined) are assayed. It can be expected that this newly obtained mitochondrial sequence will be especially useful to polarize mitochondrial data sets with which one can hope to investigate the relationships among the most species-rich class of vertebrates. The firm establishment of the phylogenetic position of the bichir awaits the collection of additional data sets, e.g., the complete mitochondrial genomes of an acipenseriform fish (e.g., sturgeon) and a cartilaginous fish.

The resolution of the bichir's phylogenetic position has bearing on the understanding of the evolutionary history, establishment of homology, and the evolution of developmental mechanisms that led from fish fins to vertebrate limbs. Pentadactyl tetrapod limbs have evolved from sarcopterygian lobed fins (e.g., Ahlberg and Millner 1994; Coates 1994). The bichir's phylogenetic position would suggest that its lobed fins might not be a shared derived trait that unites them with sarcopterygians but rather that the birchir's lobed fins had evolved independently (Figure 6). Fossil evidence (e.g., reviewed in CARROLL 1988) already indicated that fossil actinopterygian fish had endochondrial radial bones that did not extend far out into the fin. The birchir's probable phylogenetic positions as an actinopterygian would therefore argue for independent evolution of its lobed-fins from the plesiomorphic actinopterygian condition. The disputed homologies (reviewed in e.g., Patterson 1980, 1982; Ahlberg and Millner 1994; Coates 1994) for the axial elements of the pectoral appendages of fish and land vertebrates and the evolution of their development (Figure 6) will be facilitated by the firm establishment of the phylogenetic position of the bichir.

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[^1]:    Mitochondrial DNA was purified from fresh liver and kidney tissue of commercially obtained bichir (Polypterus ornaiipinnis) as previously described (ZaRDOYA et al. 1995). After homogenization, intact nuclei and cellular debris were removed by low-speed centrifugation ( $1000 \times g$ ). Mitochondria were pelleted by centrifugation at $10,000 \times g$ for 20 min and subjected to a standard alkaline lysis procedure. The isolated mtDNA was cleaved with EcoRI and HindIII restriction enzymes. Three EcoRI fragments of $4.0,3.9$ and 3.0 kb and four HindIII fragments of $3.1,1.0,0.6$, and 0.3 kb (See Figure 2 for positions of restriction sites) were cloned into pUC18 covering the entire bichir mtDNA molecule except for a stretch of 3.1 kb spanning the $C O I$ and $C O I I$ region. This

[^2]:    ${ }^{a}$ Ray-finned fish refers to the average sequence similarity between bichir against trout, carp and loach.

