



SYMPOSIUM

Grand Challenges in Comparative Tooth Biology

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Synopsis Teeth are a model system for integrating developmental genomics, functional morphology, and evolution. We are at the cusp of being able to address many open issues in comparative tooth biology and we outline several of these newly tractable and exciting research directions. Like never before, technological advances and methodological approaches are allowing us to investigate the developmental machinery of vertebrates and discover both conserved and excitingly novel mechanisms of diversification. Additionally, studies of the great diversity of soft tissues, replacement teeth, and non-trophic functions of teeth are providing new insights into dental diversity. Finally, we highlight several emerging model groups of organisms that are at the forefront of increasing our appreciation of the mechanisms underlying tooth diversification.

Introduction

Teeth provide an important phenotype for integrating across biological disciplines ranging from ecology to genomics. For instance, teeth are used to make inferences about ancient as well as recent ecologies (Purnell et al. 2007; Cuzzo et al. 2014), to understand how highly conserved structures are biomechanically modified to generate novel organismal functions (Silverman and Dunbar 1980; Parmentier et al. 2017), as well as to understand tissue development (Mitsiadis et al. 1998; Tucker and Sharpe 2004), cell (Sharpe 2001), and gene interactions (Thesleff and Sharpe 1997; Jernvall and Thesleff 2012; Jackman et al. 2013). Because all vertebrate teeth are homologous and derived from mineralized tooth-like structures present in a common early ancestor (Smith and Coates 1998; Smith 2003; Fraser and Smith 2011; Rasch et al. 2016), they are an excellent organ system for determining how multiple

levels of biological complexity have comparatively contributed to vertebrate diversification. Serially homologous teeth with different phenotypes also frequently co-occur in the same trophic apparatus (Liem 1973; Herring 1993; Vonk et al. 2008; Fraser et al. 2009; Hulsey et al. 2016; Karagic et al. Forthcoming 2020a). We can leverage these replicated structures to investigate mechanisms of tooth formation that contribute to differentiation in a single organism. Well-studied mammalian dentitions represent only a small subset of vertebrate dental diversity, so comparative studies of teeth in non-mammalian vertebrate models will likely provide the next generation of transformative insights concerning tooth diversification.

In this review, we identify research areas in comparative tooth biology that represent important challenges for the field and provide possibilities for significant future scientific advances (Table 1).

Table 1 Grand challenges in comparative vertebrate tooth biology

Developmental genomic challenges	
Deep homology of teeth	
Conserved genetic basis of tooth development	
Evolutionary developmental networks and modules	
Novel genes deployed during tooth development	
Functional morphological challenges	
Interacting functions of soft and hard dental tissues	
Polyphyodont dentitions	
Non-trophic roles and multifunctionality	
Emerging models	
Bats	
Non-mammalian amniotes	
Teleost fishes	
Chondrichthyans	

The broad categories of challenges discussed and particularly compelling topics within those categories that are highlighted in the text are listed. Additionally, we highlight several emerging model groups where addressing these challenges is likely to be highly tractable.

Furthermore, we emphasize how these synthetic areas are becoming increasingly tractable because of recent advances in technology, analytical approaches, and the comparative study of dentitions in emerging model groups. Each of these grand challenges is a call for collaborative and interdisciplinary work that integrates knowledge across fields, levels of biological organization, and incorporates studies on new lineages of vertebrates (Fig. 1). The grand challenges we outline in developmental genomics and functional morphology of dentitions should also provide insight into the need for training and educating future comparative tooth biologists. Because teeth and human health are inextricably linked (Nesse et al. 2006; Gibbons 2012; Boughner and Rolian 2015; Hovorakova et al. 2020), each challenge holds real potential for translational interactions with the field of dentistry.

Developmental genomic challenges

Developmental genomics is undergoing a revolution as technological advances make the integration of genomic and gene expression data tractable like never before. These new technologies and methodological approaches are allowing us to peel back the developmental machinery of vertebrates and expose new and exciting generalizable patterns. It is clear that many structures like teeth often share highly conserved developmental pathways with other organs, and that all vertebrate teeth are generated

using some of the same genes (Fraser et al. 2010). But, a comprehensive understanding of developmental genetics will require an appreciation of how genes interact in networks as well as determining if genes that are unique to particular lineages of vertebrates play an important role in the evolutionary development of teeth (Hulsey et al. 2016).

Deep homology

Teeth have long been recognized as being homologous to other vertebrate integumentary organs. Remarkably, Darwin (1875), long before any understanding of developmental genetics, but after encountering humans and dogs lacking both teeth and hair, wrote “The skin and the appendages of hair, feathers, hoofs, horns, and teeth, are homologous over the whole body . . .”. Over a century later, work based on comparative histology and subsequent developmental genetics has supported different levels of homology for teeth and other epithelial appendages, including scales, feathers, mammary glands, and hair (Pispa and Thesleff 2003; Dhouailly et al. 2019). On a morphological level, homology of these structures rests on early development when they all first arise via placodes, local thickened epithelial swellings overlying a mesenchymal condensate (Ørving 1967). Genetic data showing that the same genes pattern diverse epithelial appendages further supported the hypothesis of homology. For example, ectodermal dysplasia, the phenotype that likely inspired Darwin to propose homology between teeth and hair, affects teeth and hair in humans, mice, and dogs. This condition was found to result from single gene mutations (e.g., in *Ectodysplasin* [*Eda*] and its receptor [*Edar*]) (Sadier et al. 2014). Furthermore, the finding that mutations in the orthologs of these genes in distant vertebrates including reptiles and fish show that this gene regulatory network involving *Eda* and *Edar* has regulated these appendages for several hundred million years of vertebrate evolution (Kondo et al. 2001; Harris et al. 2008; Di-Poi and Milinkovitch 2016).

This homology of the different epithelial appendages raises the question of at what level of biological organization are these different vertebrate organs homologous (Abouheif et al. 1997). One favored model has involved co-option, that the gene regulatory network that evolved first to make a scale or a tooth, was subsequently repurposed at other anatomical sites (Mindell and Meyer 2001). For example, the initial observation that *Edar* in medaka fish is required for scale formation led to the proposal that this circuit was likely co-opted into reptilian

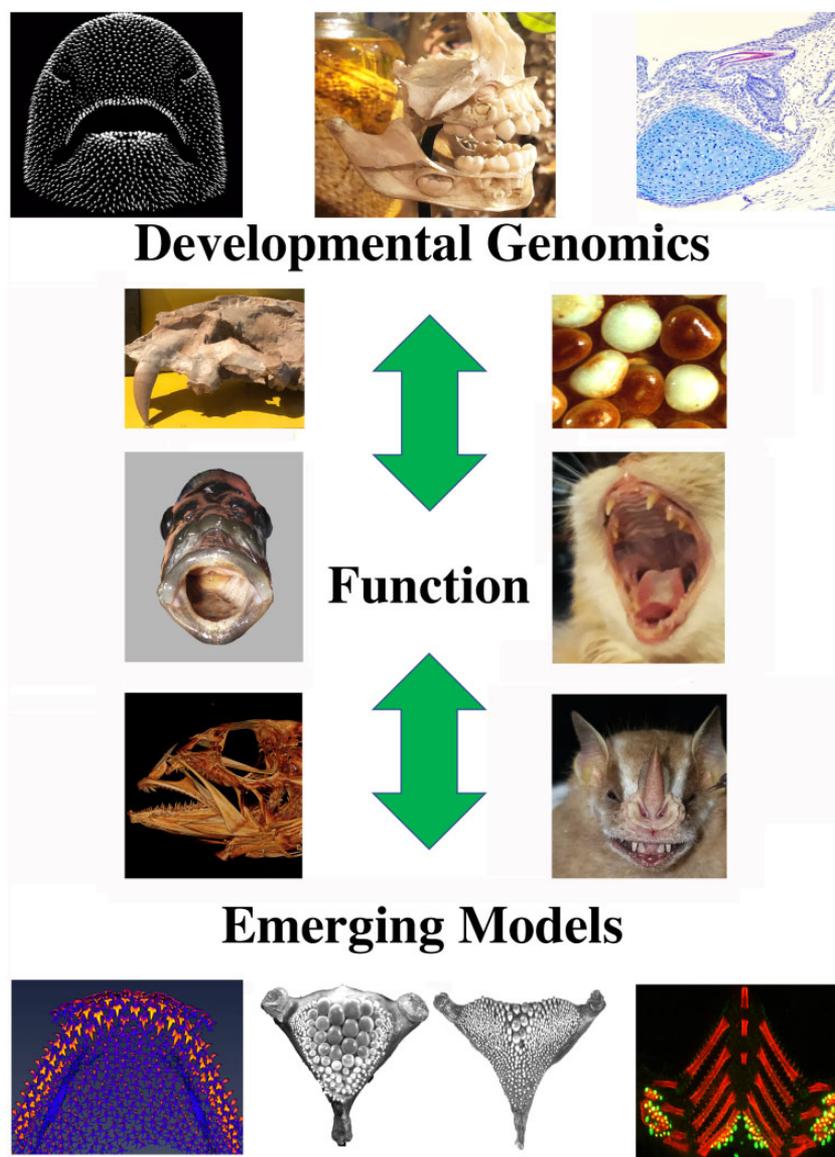


Fig. 1 The scientific interplay between developmental genomics, dental function, and emerging models of tooth diversification is highlighted visually.

scales that then evolved into mammalian hair (Sharpe 2001).

To date most developmental genetic studies of epithelial appendages have focused on the development of the initial primary organs. However, the ancestral state of tooth development is polyphyodonty (the constant regeneration of teeth throughout the lifetime of the adult animal), a condition retained in extant sharks, fish, and reptiles (Handrigan and Richman 2010; Martin et al. 2016; Rasch et al. 2016; Thiery et al. 2017; Bertin et al. 2018; Salomies et al. 2019). Since many other epithelial appendages also regenerate, one outstanding question is whether the genetic regulatory networks regulating epithelial appendage regeneration are also

shared across diverse epithelial appendages. Recently, developmental genetic studies have begun to focus on the genetic pathways regulating epithelial appendage regeneration (Wu et al. 2013; Hsu et al. 2014; Tucker and Fraser 2014; Ellis et al. 2015; Rasch et al. 2016; Aman et al. 2018). The most intensely studied regenerating epithelial appendage is mammalian hair, and decades of elegant genetic studies have revealed deep insights into the gene networks that regulate hair regeneration. This work has supported a model where organ regeneration is negatively regulated by the bone morphogenetic protein (BMP) pathway and positively regulated by the Wingless (Wnt) signaling pathway (Kobielak et al. 2007; Kandyba et al. 2013). Genetic studies in fish have

yielded some support for a model where teeth and hair regenerate using a shared genetic circuit, as the *Bmp6* gene has been implicated in negatively regulating fish tooth regeneration (Cleves et al. 2018), perhaps similar to the role of mammalian *Bmp6* in negatively regulating hair regeneration (Wu et al. 2019). Whether or not these genetic similarities extend to a larger gene regulatory network and support homologous networks governing the regeneration of teeth and other epithelial appendages remains a grand challenge for future work.

The conserved genetic basis for tooth development

Despite the incredible diversity in form, arrangement, and regenerative capacity of vertebrate dentitions, tooth development across all vertebrates is in many ways surprisingly highly conserved (Jernvall and Thesleff 2012; Tucker and Fraser 2014). Essentially, the same cells express a highly similar network of genes to coordinate the construction of vastly different teeth from fish to mammals. Conservation of this system is a rigid developmental foundation, whereby evolution has routinely modified from but retained the developmental and overall functional veracity of the vertebrate dentition (Fraser et al. 2009). Recently, genomic and developmental advances in the study of odontogenesis have led to new perspectives focused on the implications of this conservation and the discovery of novel genetic markers involved in the development and renewal process (Salomies et al. 2019; Seidel et al. 2017; Sharir et al. 2019). One such implication of this conservation is that knowledge of how distant groups, such as elasmobranchs, reptiles, and rodents, are able to maintain continuous or perpetual tooth production could highlight essential elements (i.e., cellular or genetic) that have disappeared in other groups that have reduced or have completely lost tooth regeneration and/or renewal.

The ability to make and then replace teeth is highly dependent on the activity and maintenance of the dental lamina, an epithelial sheet from where teeth originate (Smith et al. 2009; Martin et al. 2016; Rasch et al. 2016). This dynamic layer of epithelial cells is diverse in its form and function, with some vertebrate groups only able to produce a single generation of teeth, whereas others (e.g., sharks) have a near infinite supply of developing teeth (Huyseune and Sire 1998; Rasch et al. 2016). Thus, the assumption is that integrity of this cell layer is vital to further tooth production and that its degradation is related to the inability to develop further tooth generations (Popa et al. 2019). As our knowledge of the

dental lamina and its persistence among lineages improves, we can further test the role of the dental lamina in tooth replacement. For instance, sharks exhibit perpetual regeneration of the dentition and this is linked to the constant proliferative activity, growth, and maintenance of stem/progenitor sites within the entirety of the dental lamina (Smith et al. 2009; Martin et al. 2016; Rasch et al. 2016). However, in mammals that exhibit a restricted number of tooth generations, typically two (Diphyodonty), it is thought that the breakdown of the dental lamina is responsible for the restricted and limited productivity of the dentition (Buchtová et al. 2012). Additionally, several recent studies have emerged that shed a different light on the ultimate fate of the dental lamina in mammals. Even though the typical mammalian dental lamina degrades via apoptosis, some cell clusters are retained in the oral epithelium/gingiva (Buchtová et al. 2012). These clusters of remnant dental lamina, or dental rests, that have no clear function are maintained into adulthood and contain cells that share a similar stem/progenitor cell signature to the active shark dental lamina, albeit with a lower number of proliferative cells (Fraser et al. 2019).

Biologists are now taking advantage of the availability, not only of new organismal model systems (see below), but also taking note of the comparative value of these species toward a more directed understanding of the human dentition (Fraser et al. 2019). This utility of animal models in understanding the human dentition is generally based on the concept of genetic and developmental conservation. Interestingly, new evidence suggests that human tissue may have a greater regenerative capacity than previously thought, at least with some form of stimulation. The dental lamina in humans for example degrades after two generations of tooth formation and sequential molar formation, but pockets of these rested lamina cells, either stay rested, disintegrate, or become tumorigenic (Buchtová et al. 2012; Fraser et al. 2019). The trigger for dental pathologies emerging from rested epithelia is complex; however, in some cases these tumors can produce characteristic dental tissues suggesting that the fate of these cells has always been dental, i.e., of dental lamina origin (Heikinheimo et al. 2015). With knowledge of active and controlled proliferation for new teeth, i.e., in the shark model, combined with genomic information from rested and aberrant dental lamina in mammals, a grand challenge of comparative tooth biology will be to piece together the necessary ingredients for natural tooth formation in humans for future tooth generations.

Evolution of gene expression networks and modules

Tooth development has been intensively studied using morphological comparative analyses, *in vivo* and *in vitro* methods, as well as through mathematical modeling. Because of this, the developmental network giving rise to teeth is one of the best described and best understood gene regulatory networks for the development of any organ (Jernvall and Thesleff 2012; Salazar-Ciudad 2012). Importantly, these studies have demonstrated that the diversity observed in tooth morphology among mammals is shaped by the conserved developmental mechanisms that control tooth development. As a result, dental organs represent an ideal case to study how the structure of gene regulatory networks influence the evolution of phenotypes (Salazar-Ciudad and Jernvall 2010; Uller et al. 2018). Among the developmental processes proposed to influence the direction of phenotypic evolution, the modular organization of gene regulatory networks shows particular promise. This theory suggests that the arrangement of gene regulatory networks into sub-units, also sometimes called sub-modules, can influence the direction of evolution by facilitating or constraining the appearance of new phenotypes (Lipson et al. 2002; Kouvaris et al. 2017; Uller et al. 2018).

In a modular organization, some units can be more conserved than others, with conserved core units realizing essential function of the phenotype while less-conserved, peripheral sub-units being more susceptible to variation. Some recent work (reviewed in Uller et al. 2018) has theorized about the importance of developmental network topology in biasing evolution based on inferences from empirical examples. Studies mixing developmental and computational experiments have shown that teeth are patterned by semi-autonomous sub-units of gene regulatory networks or modules (Salazar-Ciudad and Jernvall 2010; Jernvall and Thesleff 2012; Lacquaniti et al. 2013). This patterning lies at the root of tooth diversity. However, there is little direct experimentally-based knowledge of how the topology of gene regulatory networks (i.e., in sub-networks) are modified in response to environmental selection (Dumont et al. 2012; Usui and Tokita 2018).

Teeth are serially homologous structures that vary greatly in number and shape across mammals. Evolutionary changes in molar size, shape, and in the presence, size, and shape of individual cusps (Fig. 2) are all critical for increasing feeding performance within particular food types or enabling dietary transitions across food sources (Hunter and

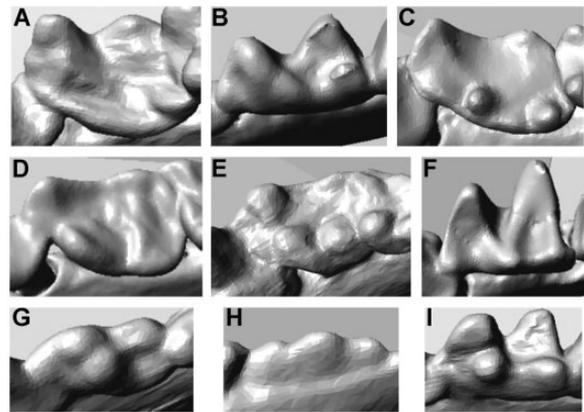


Fig. 2 Bat first molar diversity: from left to right, up to bottom; CT scan reconstructions of molars in the bats **A)** *Uroderma bilobatum*, **B)** *Macrotus waterhousii*, **C)** *Artibeus phaeotis*, **D)** *Brachyphylla pumila*, **E)** *Centurio senex*, **F)** *Noctilio leporinus*, **G)** *Sturnira lilium*, **H)** *Desmodus rotundus*, and **I)** *Trachops cirrhosus*.

Jernvall 1995; Santana et al. 2011a). For instance, comparative studies have demonstrated that the evolution of specific molar morphologies, including their size and shape, can be highly labile and evolve independently of other tooth traits. However, experiments regarding the influence of the modular structure of gene regulatory networks on phenotypic evolution have remained limited to model species such as mice and to computational predictions for other species (Häära et al. 2012; O'Connell et al. 2012; Salazar-Ciudad 2012; Harjunmaa et al. 2014). As teeth in general and mammalian molars in particular exhibit a modular organization, both phenotypically and in their gene regulatory networks, teeth constitute a powerful model to study these questions. With increasing access to new species thanks to next-generation sequencing and reverse genetic techniques such as CRISPR/Cas9, we have arrived at an exciting time when we can test hypotheses about network modularity in a comparative framework. Teeth represent an excellent model system to study how the inherent structure of gene regulatory networks influence variation and the evolution of actual and potential morphospaces. Studies of the evolution and development of teeth will help to advance the extended evolutionary synthesis by potentially finding new paradigms for the mechanisms underlying trait evolution in general.

Novel genes deployed during tooth development

As discussed above, extensive knowledge exists concerning the developmental mechanisms involved in odontogenesis in mammalian model systems especially in mice (Tucker and Sharpe 2004; Thesleff 2006). It has become axiomatic that many

developmental programs involved in tooth development are conserved across all vertebrates. Even highly derived phenotypes such as constantly growing mouse incisors (Wang et al. 2007), pufferfish beaks (Fraser et al. 2012), the crushing dentition of rays (Underwood et al. 2015), or venom-delivering fangs in snakes all follow the same developmental blueprint (Vonk et al. 2008; Landova et al. 2020). However, what could differ extensively during the development of teeth in disparate vertebrates are genes further downstream of signaling pathways such as genes involved in tooth mineralization that can be very lineage specific (Kawasaki et al. 2004; Kawasaki and Weiss 2008; Kawasaki 2009). Studies on non-model organisms have begun to show evidence that there could be substantial developmental divergence among vertebrates during tooth formation (Kawasaki et al. 2004). Additionally, there is great diversity regarding dental phenotypes in vertebrate lineages, especially in teleost fish (Liem 1973; Mehta and Wainwright 2007; Fraser et al. 2012; Berkovitz and Shellis 2018). To understand the source of this incredible diversity, a more complete approach to the genomics of tooth formation will be required and genes, other than those that have been shown to have highly conserved roles like Wnts or Efgs, will need to be studied in more detail.

For instance, investigations into the secretory calcium-binding phosphoprotein (*scpp*) gene family illustrate how divergent genetic mechanisms involving novel genes can generate diversity during tooth formation (Kawasaki et al. 2005). *Scpps* are genes present in all vertebrates and are involved in processes such as tooth mineralization (Kawasaki and Weiss 2008). All members of the *scpp*-family, including prominent tooth genes like *amelogenin* or *enamelin*, evolved from *sparc* which was present in the common ancestor of vertebrates and have since diverged independently in various lineages (Kawasaki 2009). Comparing teleosts and mammals, considerable differences in terms of gene family size, homolog sequences, and potentially their respective functions have been reported (Van de Peer et al. 2003; Hoegg and Meyer 2005). The *scpp* gene family illustrates how conserved structures and processes can have considerable variation in their developmental genetic basis and comparable variation could potentially underlie much odontogenetic diversity across vertebrates. Considering the immense diversity of trophic adaptations exhibited by teleosts and other vertebrates (Liem 1973; Eastman 1977; Hulsey 2006; Wang et al. 2007; Vonk et al. 2008; Mehta and Wainwright 2007; Fraser et al. 2009,

2012;), it is very likely that a large number of genes involved in odontogenesis remain unknown.

Novel tooth genes, here defined as genes not present in or not known from mammalian odontogenesis, could readily have contributed to the dental diversity seen in groups like teleost fishes. Following the whole genome duplication shared by most teleosts, many novel genes with divergent functions likely arose by sub- or neofunctionalization (Ohno 1970; Van de Peer et al. 2003), as illustrated by the *SCPP* gene family. Additionally, there has likely been the birth of additional novel genes from non-coding regions that have evolved to produce functional proteins. This type of gene birth was once regarded as likely to be a rare event, but its frequency and potential role in adaptation has recently been appreciated to be more common and evolutionarily important (Chen et al. 2010; Wu et al. 2011; Wu and Zhang 2013; McLysaght and Guerzoni 2015). Although the identification of such novel tooth genes remains challenging, novel genes associated with the repeated evolution of larger teeth have for instance been identified in cichlid fishes (Karagic et al. 2020a). Now that whole genomes are rapidly becoming available for many species, comparative genomic approaches will allow us to shed light on the presence of many previously unknown paralogs or orthologs as well as completely novel genes (Kawasaki et al. 2005). In general, these genes will need to be further validated regarding their role during tooth formation and for recently evolved genes, it will often be exceptionally difficult to infer gene functions due to the lack of comparative approaches. However, high-throughput transcriptomic approaches that facilitate examinations of the expression of all genes in a certain tissue, e.g., tooth buds, odontoblasts, or even single cells, could provide a much more detailed picture of what genes are active during odontogenesis and facilitate rigorous comparisons among taxa. Powerful new sequencing technologies coupled with the increased use of non-model organisms will allow us to rise to the challenge of unraveling how important previously undocumented and novel genes are to the diversity of vertebrate tooth formation.

Functional challenges

Teeth bear the stamp of their primary function in capturing and processing prey however, our understanding of the extensive functional diversity of dental structures is far from complete. Dentitions are more than a battery of hard structures and the

function of soft tissues in vertebrate dentitions are receiving greater appreciation. The extensive focus on mammalian dentitions has come at the expense of understanding the role that frequent tooth replacement likely plays in the dentition of most vertebrates, but this is changing. Additionally, there is often a match between perceived tooth function in feeding and shape, but this is not always true. For instance, because teeth are often sexually dimorphic and used for many non-trophic tasks, feeding should not remain the only possible *de facto* explanation for dental adaptation and diversity.

Interacting functions of soft and hard dental tissues

Teeth are the hardest part of the dental battery and have driven research on dentition for 200 years (Owen 1845; Green et al. 2019). However, the dental papilla, dental ligament, and to a lesser extent the tooth pulp have all been shown to play crucial roles in the development and evolution of teeth (Huysseune and Witten 2008; Denes et al. 2019). However, a grand challenge remains in understanding the function of dental soft tissues, particularly as they relate to polyphyodont dentitions.

The periodontal ligament provides an example of a multifunctional soft tissue that in mammals serves to dampen transient impacts, facilitates post-eruptive tooth movement, and plays an essential role during chewing (McCormack et al. 2014; Underwood et al. 2016). This ligament is also present in crocodylians, that are polyphyodont, providing the potential for comparative analyses to understand the functional importance of this structure (Bertin et al. 2018). This might be especially valuable as it is unclear if transient loads matter in the context of a polyphyodont dentition outside of mammals. Also, in the absence of a periodontal ligament, it is unclear how polyphyodont teeth manage variable loading conditions while simultaneously maximizing tooth performance. Polyphyodont teeth can be attached to the bone by collagen (e.g., *Iguana* and *Salmo*) (Huysseune and Witten 2008) or through ankylosis (e.g., *Chameleo* and *Pomatomus*) (Bemis et al. 2005), a process in which the tooth fuses to the jaw and loses its connection to surrounding soft tissue. Flexibility of attachment modalities in the evolution of polyphyodont dentitions may have commonly released functional constraints imposed by the periodontal ligament.

Not only do polyphyodont dentitions vary in their attachment to the jaw, they also vary in another way that has functional implications—the developmental trajectory from a germ to a fully mature tooth

(Fraser et al. 2013; Tucker and Fraser 2014). Tooth replacement patterns vary from completely extraosseous replacement in which the teeth develop in the oral epithelium adjacent the jaw to intraosseous patterns in which the tooth germ migrates into the bone to complete development (Trapani 2001; Bertin et al. 2018). The migration of a tooth into the jaw bone results in substantial remodeling that affects not only the structural integrity of the jaw, but also the replacement tooth size and shape (Bemis et al. 2005; Witten and Huysseune 2009; Bertin et al. 2018). These biological realities raise many questions such as: are replacement modalities contingent on the structural mechanics of the jaw? How does the dental battery change functionally as teeth of different shapes are ankylosed in new positions? Can a tooth that develops outside of the jaw be as robust as one that develops inside a bony crypt? We need histological and functional studies of tooth development from a diversity of species to answer these questions.

Teeth are characterized by having a center of vascularized pulp and the proportion of the pulp cavity relative to the dentine and enamel is highly variable across species (e.g., Huysseune and Sire 1998). The biomechanics of teeth, like any organ, are governed by structural and material properties (Anderson and LaBarbera 2008). But, how much of tooth function is determined by the pulp cavity? Take for example two cases of “fang-like” teeth in fishes: the hollow fangs of *Alepisaurus ferox* and robust fangs of *Ophiodont elongatus*. Both have small conical teeth interspersed with larger fangs (Gallo and Levin 2016). The diets are different: *A. ferox* consumes gelatinous prey (Kubota and Uyeno 1970) while *O. elongatus* is a piscivore. This is one of many examples where anatomical descriptors such as “fang” or “conical” provide limited information to link tooth form and function. Perhaps the structure and/or material of the teeth of *A. ferox* are incapable of resisting abrasive forces generated during puncture. Quantifying proportions of pulp, dentine, and enamel is one simple way that could be used to separate these and other dentitions. A better understanding of the performance of soft tissue will help us answer questions of tooth complexity within taxonomic and functional groups from new perspectives.

Soft tissue is likely as variable functionally and morphologically as the hard parts of the dental battery, and could be even more important in polyphyodont dentition when compared with vertebrates with diphyodont dentitions. Evaluating soft tissue with respect to function, instead of as an incidental necessity of tooth development, will reveal patterns in polyphyodont dentitions that

have otherwise been obscured by the focus on the hard elements of vertebrate dentitions.

Distinct functions of polyphyodont dentitions

The teeth of some fishes, when considered as a battery, are so unusual as to defy hypotheses of function (Fig. 3). The hard tissue of the dental battery has been a fertile ground for research, with particular attention to the physical and mathematical modeling of the relationship between form and function (Herring 1993; Evans and Sanson 2006; Crofts and Summers 2014; Freeman and Lemen 2007; Ramsay and Wilga 2007; Anderson and LaBarbera 2008; Anderson 2009; Whitenack and Motta 2010; Santana et al. 2011a; Evans 2013; Smits and Evans 2012; Berthaume et al. 2014). The biomedically relevant and extensive focus on mammalian teeth gives us a biased view that now provides a grand opportunity for understanding the performance consequences of possessing a polyphyodont dentition rather than one with a single replacement cycle (Huysseune and Sire 1998). We have substantial gaps in our understanding of polyphyodont dentitions including unexplained patterns of tooth size, a variety of tooth replacement modes, and tooth shapes that have no intuitive function.

For instance, the diversity of dental batteries in fishes leads to speculation that continuous replacement of teeth makes possible tooth shapes, positions, and orientations that seem precarious (Fig. 3A). For example, in some deep-sea fishes (e.g., *Anoplogaster*, Fig. 3B) the teeth are so slender and tall that it is hard to imagine that they can be used to penetrate anything of substance. Another species (e.g., *Omosudis*) has alternating groups of large and small blade-like teeth. Yet another (e.g., *Anopterus*) has blade-like teeth pointing anteriorly. What drives this diversity? Could it be driven by the lack of constraint inherent in continuous, cheap replacement teeth?

In diphyodont animals, replacement teeth default to a homologous position, but in many polyphyodont dentitions replacement teeth are not homologous to the functional tooth they replace (i.e., a bigger tooth may replace a small tooth). Gaengler (2000) proposed the question “can a canine be incisiform”? Highlighting the deeper question: is the function of a tooth more determined by shape or by position? If tooth shape dominates function, then an incisor in the place of a molar should still function as an incisor. This predicament is further muddled when we consider the functions of similarly shaped teeth (e.g., fang versus cones) (Olson 2017).

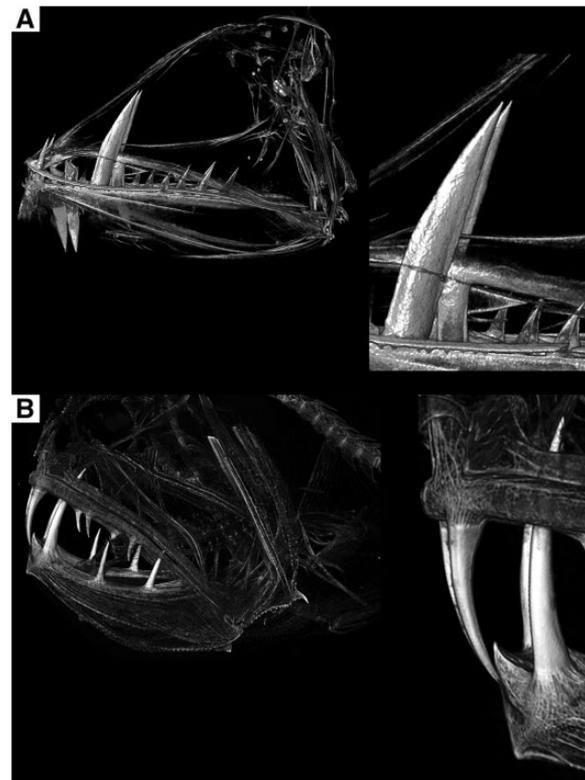


Fig. 3 A) CT image of *Omosudis lowii*. Note the enlarged fangs at the front of the dentary followed by smaller conical teeth posteriorly. B) CT image of *Anoplogaster cornuta* with large, curved fangs extending from both the upper and lower jaws.

Mihalitsis and Bellwood (2019) addressed the divergence in form and function by recognizing that position and orientation of conical teeth changes function. This is because similarly shaped teeth at opposite ends of the jaw will necessarily have different amounts of forces acting on them. Berthaume et al. (2013) showed that it took a combination of dull and sharp cusps to create an ideal grinding surface, rather than a single tooth or tooth type. Whitenack and Motta (2010) found that many different shark tooth morphologies were functionally equivalent. Much research has focused on geometric ways to create ideal puncturing, grinding, or slicing tools (Anderson 2009; Whitenack and Motta 2010; Berthaume et al. 2013; Crofts and Summers 2014; Anderson et al. 2016). But, the functional implications of a single tooth could also change when placed in the context of the entire dentition.

Homodonty and heterodonty only begin to describe the entire battery by associating the regionalization of tooth types with a regionalization in function (Keene 1991; Cohen et al. Forthcoming 2020). These terms are vague and provide little quantitative understanding of a tooth or dental function. Even when we incorporate phylogenetic,

developmental, or ontogenetic constraints, these diphyodont-centric terms provide no space for constraints, limitations, or plasticity provided by a polyphyodont dentition (Karagic et al. 2020b, in review). The enormous range in polyphyodonty asks us to reconsider what constraints the function of a tooth has in the context of a functioning dental battery. If polyphyodonty is an ancestral trait, then what does heterodonty and homodonty truly represent? Is a homodont dentition one where all of the teeth, despite shape or size variation, perform in the same way? Quantifying tooth shape in terms of function allows us to explore teeth relative to how they interact with a prey item. By incorporating geometric constraints back into our understanding of the dental battery, we can begin to quantify tooth complexity, integration, and morphology of polyphyodonts in new functional and evolutionarily meaningful ways.

Non-trophic roles and multifunctionality

Teeth provide an exceptionally interesting phenotype for examining the interplay between natural and sexual selection (Darwin 1871; Katsikaros and Shine 1997; Gorman and Hulsey 2020, in review). For instance, many tooth sexual dimorphisms have been shown to be associated with divergent male and female feeding ecologies (Dayan et al. 1989). Yet, although tooth sexual dimorphism could often foster trophic differences, it might also commonly reflect other functions of teeth that have diverged between males and females that are not related to subduing or processing prey (Randau et al. 2013). For instance, teeth are often used in aggressive displays related to dominance hierarchies in groups such as primates and carnivores (Harvey et al. 2009). Many groups such as elephants, vampire deer, and the narwhal also use their teeth specifically for intraspecific contests often involving male–male combat (Silverman and Dunbar 1980). Teeth could also commonly be used for marking territories in ungulates and other territorial organisms. Defense against predators, especially when one sex is more susceptible to predation, could likewise be a cause of tooth dimorphism (Cowlisa 1994; Ovsyanikov 1995). Teeth could often promote ecological divergence and specialization that extends beyond the direct functional consequences of trophic needs.

The extensive sexual dimorphism in teeth also highlights the multi-functionality of teeth (Gorman and Hulsey 2020, in review). For instance, the grinding or scraping of teeth that are otherwise specialized for certain foods can be used for intraspecific vocalizations and mating calls. Mammals such as guinea

pigs can engage in teeth chattering and many teleost fish species putatively use their pharyngeal teeth for inter- and intra-specific communication (Parmentier et al. 2017). Additionally, the morphology of teeth in the front of the oral cavity likely influences non-tooth vocalizations of many mammals such as dolphins and bats (Perrin et al. 2011). Additionally, teeth could commonly be used for grasping mates during copulation. This is likely the case for the sexually dimorphic teeth found in groups such as stingrays and other elasmobranchs (Kajiura and Tricas 1996). Many teeth have also been modified into sexual ornaments for attracting the opposite sex (Hendrie and Brewer 2012). Groups of primates and other mammals also likely use teeth during grooming (Rose et al. 1981). A number of groups such as crocodylians, cardinal fishes, and cichlid fishes also brood their offspring in their mouths (Hulsey 2009). Teeth could therefore be modified to enhance this mouth brooding or maternal care. There could commonly be substantial interactions between natural and sexual selection in structuring the evolution of teeth, and as we increase our ability to evaluate multiple functionality, teeth will continue to be an excellent trait to highlight the relative importance of these two forces.

Because teeth do generally have extensive roles in trophic ecology, understanding the mechanisms related to the switch to using teeth for novel functional tasks associated with sexual dimorphism might also be especially tractable (Gorman and Hulsey 2020, in review). There are several vertebrate groups for which sex-specific tooth modifications have been suggested to confer highly novel functions. For instance, male narwhals are thought to use their enlarged single tusks as salinity gauges (Nweeia et al. 2014). Many vertebrates can also use their teeth for novel locomotory functions. For instance, walruses use their tusks for hauling themselves onto ice-shelves (Fay 1982). Many fishes such as gobies and catfishes also use their sucker mouths for holding station in flow or moving through torrential environments such as waterfalls and teeth could be critical here (Geerinckx and De Kegel 2014). Groups such as elephants and rodents also use enlarged teeth for forcing or digging their way through complex environments (McIntosh and Cox 2019). In light of our extensive understanding of the developmental genetics and evolutionary history of teeth, non-trophic dental functions could provide challenging but highly tractable systems for examining the integrative origin of novel evolutionary abilities.

Emerging animal models

To better understand the developmental genomics and function of all vertebrate tooth diversity, we obviously need to incorporate studies of many vertebrate groups. But, comparative biologists have honed in on a few groups such as bats, reptiles, rapidly diversifying teleosts, and chondrichthyans that show particular promise in shedding new light onto the fundamental mechanisms governing the developmental genomic, functional divergence, and evolution of vertebrate tooth diversity.

Bats as eco–evo–devo models

Thanks to the rise of non-model systems, it is now possible to investigate evolutionary morphological variation at the genomic, developmental, and phenotypic levels *in situ*. In this respect, bats (Chiroptera) represent an iconic example: currently with 1411 described species (<https://batnames.org>), and an incredible craniofacial diversity (Usui and Tokita 2018), bats are now considered a key evo–devo group to study craniofacial variation. Following the acquisition of powered flight, bats underwent a major adaptive radiation into a range of dietary niches that encompass nearly all possible mammalian diets, (i.e., fruit, nectar and pollen, leaves, seeds, arthropods, small vertebrates, fish, and even blood) (Dumont *et al.* 2012). To be able to exploit this extensive range of diets, bats have evolved a wide diversity of tooth morphologies, including variation in width, length, height, and (in the case of molars) cusps, that has been intensively studied in the last 30 years (Freeman 1988, 1992; Santana and Dumont 2009; Santana *et al.* 2011a, 2011b; Dumont *et al.* 2012; Santana and Cheung 2016). Bats exhibit a classic mammalian tribosphenic molar design (Freeman 1992): a first molar (m1) with five cusps that is derived from the ancestral mammal condition by loss of the hypoconulid. Additionally, some mechanisms have been proposed to explain their craniofacial variation in relationship to diet (Arbour *et al.* 2019; Camacho *et al.* 2019). Moreover, they represent a good model to study the evolution and the developmental differences between tooth classes, as some species exhibit molariform premolars or reduced canines (Crompton and Hiimae 1969; Freeman 1992) as well as other important variations of their dental formula between species (Freeman 1992; Giannini and Simmons 2007). Finally, because they are diphyodonts, bats are also a very good model for the study of tooth replacement (Popa *et al.* 2016). To date, the developmental mechanisms governing the dental morphology in non-model, wild mammal

species have been only partially understood, due to difficulties in obtaining embryonic materials for analyses or having genomes available. Thanks to new techniques, access to museum specimens (Hedrick *et al.* 2018), field caught pregnant bats, possibilities offered by *in vitro* culture for teeth, and the bat1K genome consortium, we now have access to unparalleled eco–evo–devo resources to study dental evolution in this most species-rich lineage of mammals. Together, these characteristics make bats an outstanding model to study tooth evolution in a comparative framework and to address other big questions involving such topics as tooth replacement and bioengineering *in vitro* synthesis of teeth.

Non-mammalian amniotes

Several lineages of what are generally referred to as reptiles including crocodylians, lizards, and snakes also provide exciting emerging models for tooth biology. Dental developmental genetics and functional morphology in these non-mammalian amniotes will continue to provide critical insight into the evolution of mammalian dentitions. Mammalian traits such as extensive morphological heterodonty, enlarged tooth cusps, occlusion, and reduction of tooth replacement can all be placed in a more robust historical context as we learn more about reptilian teeth (Berkovitz and Shellis 2018). For instance, a number of lineages like snakes and agamid lizards have highly heterodont dentitions with extensive shape diversity of individual teeth that form their dental arcades (Cooper *et al.* 2009; Landova *et al.* 2020). This within individual variability in teeth likely has facilitated a high degree of feeding specialization (Dumont *et al.* 2012). Tooth replacement has also likely been convergently reduced in some of these amniote lineages facilitating comparative studies (Buchtová *et al.* 2013). Likewise, crocodylians share several aspects of tooth attachment and replacement with mammals that could be functionally and genetically investigated (Wu *et al.* 2013; Bertin *et al.* 2018). An enhanced understanding of the functional morphology of extant amniotes will also provide greater insight into fossil groups especially those unique to the extinct ancestral lineages leading to mammals (Berkovitz and Shellis 2018). Further examinations of the diversity of tooth attachment, implantation, and replacement in extant and extinct reptiles will also continue to provide understanding into the ancestral condition of these fundamental dental traits in mammals and other amniote trends in tooth diversification (Bertin *et al.* 2018).

Reptile dentitions will also continue to shed light on the genetic mechanisms generating dental complexity. The deep homology of epithelial appendages, including teeth, will continue to benefit from studies of reptiles (Di-Poi and Milinkovitch 2016). Additionally, the degree to which developmental processes such as morphogenesis and replacement are genetically decoupled can be readily explored in reptiles (Handrigan and Richman 2010; Salomies et al. 2019). What developmental pathways are responsible for the continued maintenance of the dental lamina and polyphyodont replacement patterns will continue to provide evolutionarily relevant comparisons to the mammalian condition (Whitlock and Richman 2013). The genes recruited in the formation of highly novel tooth types such as snake fangs will further provide insight into the origin of dental novelty (Vonk et al. 2008; Landova et al. 2020). As developmental genomic tools become increasingly available in non-model reptiles that can be investigated experimentally, we will gain a greater understanding of all amniote dentitions including our own.

Teleost fishes

Teleost fishes provide a rich evolutionary context for the investigation of the mechanisms generating dental divergence because of the numerous axes along which their teeth have diverged phenotypically and presumably developmentally (Hulsey et al. 2016). In emerging model groups like sticklebacks, pufferfish, and cichlids, the study of how vertebrate teeth can diverge among species into phenotypically novel traits is especially evolutionarily and experimentally tractable. These species can often be brought into the lab, hybridized to study the genetic basis of tooth divergence, and now can frequently be readily manipulated using reverse genetic techniques such as CRISPR (Hulsey et al. 2017; Cleves et al. 2018; Kratochwil et al. 2018).

Research on these fishes also address another problem with using more traditional models like the mouse and its dental developmental network as a standard for all vertebrate teeth which is that unlike both humans and cichlid fishes, mice do not replace their teeth (Fraser et al. 2004). Therefore, we know relatively little about whether the genes responsible for phenotypic differentiation of vertebrate replacement teeth are generally the same genes utilized in the formation of the initial dentition (Fraser et al. 2004, 2012, 2013; Handrigan and Richman 2010). Thus, there could be substantial differences in the genes generating replacement teeth

(Streelman et al. 2003; Schneider and Meyer 2017). Importantly, unlike mammals that replace their teeth at most a single time, cichlids and most teleost fishes can replace their teeth once approximately every 100 days repeatedly throughout their life (Huysseune and Sire 1998; Streelman et al. 2003). Much of the phenotypic diversity in the teleost dentition is also set up during the time between when tooth replacement begins and the onset of reproductive activity (Ellis et al. 2015). Therefore, teleost fish offer a system that could be used to determine what genes are conserved not only during initial vertebrate tooth formation but also what genes are expressed as these structures are replaced and differentiate phenotypically into adult dentitions.

The ever-increasing availability of genomic resources is now making it feasible to conduct comparative genomic analyses, extensively manipulate gene expression and to perform functional assays of gene networks in structures such as the toothed oral and pharyngeal jaws of cichlids and sticklebacks (Hulsey et al. 2016). The exploration of regulatory mechanisms like 3'-UTRs and microRNAs that govern important aspects of all vertebrate tooth development are also receiving increased focus in teleost fishes (Xiong et al. 2018, 2019; Franchini et al. 2019). Coupling these comparative analyses with experimental approaches and modeling of the potential interactions among genes will further allow us to test the distinctiveness of individual dental modules. Teleost dentitions will continue to provide a potent system in which to examine tooth function, development, and evolution.

Chondrichthyans

The evolution of teeth was a key event in vertebrate evolution and was linked to a greater capacity for feeding as different tooth morphologies were able to facilitate distinct feeding strategies. In fact, the predatory lifestyle that arose early in gnathostome evolution depended not only on the evolution of the jaw, but also of hard tissues such as bones and teeth (Owen 1845). Along with this, patterning or organization of teeth on the jaw sets up the functional dentition, while tooth replacement ensures new teeth are readily available at the oral surface. Coordinating all these factors across the jaw requires precise positioning of new teeth and is perhaps most apparent in the vertebrate group Chondrichthyes (sharks, rays, and chimaeroids). For example, even in the very different dentitions of sharks and rays, there are similar patterns of tooth addition,

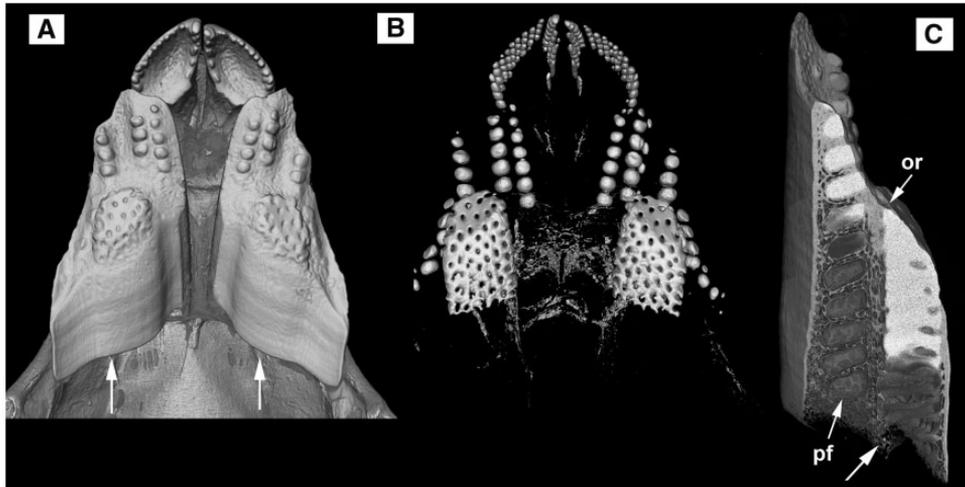


Fig. 4 *Harriotta raleighana* (Chimaeridae; Chimaeroidei; Holocephali), upper dentition, micro-CT scan. **A**) Skull and anterior and posterior upper jaw dental plate. **B**) Rendered scan to show mineralized dentine structures, including columns of ovoids and tritons. **C**) Virtual section through lower jaw dental plate showing mineralized ovoids and tritons, and surrounding trabecular dentine of lower mineral density. Tissue growth occurs at the aboral surface (opposite to the oral surface, or), including organized, preformed spaces within the trabecular dentine for ovoids and tritons. From Smith et al. (2019, figs. 2D and 16G and H).

positioning along the jaw, and replacement such that commonalities in these groups (known as Elasmobranchii) can be identified (Underwood et al. 2015, 2016).

By comparison, the third major group of chondrichthyans, the chimaeroids (=holocephalans), shows a very different type of dentition (Patterson 1965; Didier et al. 1994; Stahl 1999) involving broad dental plates in the upper and lower jaws. There is no indication of replacement teeth forming at any time during the development of these plates (e.g., Smith et al. 2019), in any of the holocephalan families (Callorhynchidae, Chimaeridae, Rhinochimaeridae). However, some researchers have suggested that different parts of these plates represent original tooth germs (Schauinsland 1903; Kemp 1984; Didier et al. 1994). There are nevertheless some intriguing similarities to replacing and patterning chondrichthyan dentitions in the holocephalans. For example, dental plate tissues are clearly replaced at the postero-lingual margin of the plate, in close association with the base of the cartilage jaw (Smith et al. 2019; Fig. 4A, C), in a position similar to the dental lamina in elasmobranchs, and the tooth-producing tissues located there (Smith et al. 2009; Underwood et al. 2015; Martin et al. 2016). Also, multiple mineralized dentine elements that form part of the dental plate in the family Chimaeridae show patterning within the dental plate (Fig. 4B), including preformed spaces within the surrounding less mineralized dentine (Fig. 4C). How these spaces are organized and involve gene regulatory networks related to tooth development in

chondrichthyans and bony fishes (e.g., Fraser et al. 2009; Rasch et al. 2016) will be an interesting area for future research.

How these plates evolved is also uncertain because most fossil relatives are very similar to the living families (Patterson 1965; Stahl 1999; Johanson et al. Forthcoming 2020a). However, more distantly related taxa and new phylogenetic analyses (Coates et al. 2017, 2018) are providing a clearer picture, along with computed tomography (CT) scanning, which allows us to investigate the patterning, organization on the jaw, and histology of these dentitions in greater detail. One of the most interesting of these fossil relatives is *Helodus simplex*. *Helodus* was first described in the mid-1800s (Agassiz 1833–1843), and like many chondrichthyan fossils, was known from isolated teeth that had fallen apart after death. Rarer, more complete specimens (Moy-Thomas 1939) demonstrate that these teeth were joined to form a whorl-like structure (a common feature of fossil chondrichthyans), and that substantial fusion has occurred among the dental elements. This fusion creates a more plate-like structure, but later in development—initially these teeth are more separated in the tooth whorl (Johanson et al. Forthcoming 2020b). *Helodus* therefore provides a mechanism for holocephalan plate development, while new phylogenies resolve a range of fossil taxa with broadly similar tooth whorls as holocephalan relatives (e.g., Cladoselache; Johanson et al. Forthcoming 2020b), suggesting that a progressive loss of tooth identity was characteristic of the evolution of this group and led to the formation of a

novel dental morphology. The combination of fossil material with extant diversity will continue to be an important challenge that helps us to better understand not only what vertebrate dental diversity has existed but also how it originated.

Conclusion

In this article, we have highlighted the importance of grand challenges in tooth biology. It is clear that it will take interdisciplinary research to make real advances in this critical area of organismal biology. This process needs to include the training of new scientists in interdisciplinary fields that are likely to provide important insights and possible solutions to major problems. We also need to identify the tools that we require to make these advances. We must stimulate, encourage, and train comparative biologist who will provide us with the novel approaches necessary to open future horizons in tooth biology. Importantly, the grand challenges that we have identified are not intended to be exhaustive or to limit the focus on other important issues within comparative tooth biology. Nevertheless, frequent reassessment of our grand challenges with a focus on the future will allow us to highlight the advantages of comparative knowledge for those examining the different levels of biological organization that contribute to tooth diversity.

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References

Abouheif E, Akam M, Dickinson WJ, Holland PW, Meyer A, Patel NH, Raff RA, Roth VL, Wray GA. 1997. Homology and developmental genes. *Trends Genet* 13:432–3.

- Agassiz L. 1833–1843. *Recherches sur les poissons fossiles*. Vol. 3. Neuchâtel: Petitpierre.
- Aman AJ, Fulbright AN, Parichy DM. 2018. Wnt/ β -catenin regulates an ancient signaling network during zebrafish scale development. *Elife* 7:e37001.
- Anderson P. 2009. The effects of trapping and blade angle of notched dentitions on fracture of biological tissues. *J Exp Biol* 212:3627–32.
- Anderson PSL, LaBarbera M. 2008. Functional consequences of tooth design: effects of blade shape on energetics of cutting. *J Exp Biol* 211:3619–26.
- Anderson PSL, LaCosse J, Pankow M. 2016. Point of impact: the effects of size and speed on puncture mechanics. *Interface Focus* 6:20150111.
- Arbour JH, Curtis AA, Santana SE. 2019. Signatures of echolocation and dietary ecology in the adaptive evolution of skull shape in bats. *Nat Commun* 10:2036.
- Bemis WE, Giuliano A, Mcguire B. 2005. Structure, attachment, replacement and growth of teeth in bluefish, *Pomatomus saltatrix*, a teleost with deeply socketed teeth. *Zoology* 108:317–27.
- Berkovitz BKB, Shellis RP. 2018. *The teeth of non-mammalian vertebrates*. Cambridge (MA): Academic Press.
- Berthaume MA, Dumont ER, Godfrey LR, Grosse IR. 2013. How does tooth cusp radius of curvature affect brittle food item processing? *J R Soc Interface* 10:20130240.
- Berthaume MA, Dumont ER, Godfrey LR, Grosse IR. 2014. The effects of relative food item size on optimal tooth cusp sharpness during brittle food item processing. *J R Soc Interface* 11:20140965.
- Bertin TJC, Thivichon-Prince B, LeBlanc ARH, Caldwell MW, Viriot L. 2018. Current perspectives on tooth implantation, attachment, and replacement in Amniota. *Front Physiol* 9:1–20.
- Boughner JC, Rolian C. 2015. *Developmental approaches to human evolution*. New York (NY): Wiley Blackwell.
- Buchtová M, Stembírek J, Glocová K, Matalová E, Tucker AS. 2012. Early regression of the dental lamina underlies the development of diphyodont dentitions. *J Dent Res* 91:491–8.
- Buchtová M, Zahradníček O, Balková S, Tucker AS. 2013. Odontogenesis in the veiled chameleon (*Chamaeleo calyptratus*). *Arch Oral Biol* 58:118–33.
- Camacho J, Heyde A, Bhullar BS, Haelewaters D, Simmons NB, Abzhanov A. 2019. Peramorphosis, an evolutionary developmental mechanism in neotropical bat skull diversity. *Dev Dyn* 248:1129–43.
- Chen S, Zhang YE, Long M. 2010. New genes in *Drosophila* quickly become essential. *Science* 330:1682–5.
- Cleves PA, Hart JC, Agoglia RM, Jimenez MT, Erickson PA, Gai L, Miller CT. 2018. An intronic enhancer of *Bmp6* underlies evolved tooth gain in sticklebacks. *PLoS Genet* 14:e1007449.
- Coates MI, Gess RW, Finarelli JA, Criswell KE, Tietjen K. 2017. A symmoriiform chondrichthyan braincase and the origin of chimaeroid fishes. *Nature* 541:208–11.
- Coates MI, Finarelli JA, Sansom IJ, Andreev PS, Criswell KE, Tietjen K, Rivers ML, La Riviere PJ. 2018. An early chondrichthyan and the evolutionary assembly of a shark body plan. *Proc R Soc Biol* 285:20172418.
- Cohen KE, Weller HI, Westneat MW, Summers AP. Forthcoming 2020. A demonstration of functional

- homodonty in *Halichoeres* wrasses—mapping continuous traits. *Integr Comp Biol*.
- Cooper JS, Poole DFG, Lawson R. 2009. The dentition of agamid lizards with special reference to tooth replacement. *J Zool* 162:85–98.
- Cowlis G. 1994. Vulnerability to predation in baboon populations. *Behavior* 131:293–304.
- Crofts SB, Summers AP. 2014. How to best smash a snail: the effect of tooth shape on crushing load. *J Roy Soc Inter* 11:20131053.
- Crofts SB. 2015. The functional morphology of hard-prey crushing teeth [Ph.D. dissertation]. [Washington (DC)]: University of Washington.
- Crompton AW, Hiiemae K. 1969. How mammalian molar teeth work. *Discovery* 5:23–4.
- Cuozzo FP, Head BR, Sauter ML, Ungar PS, O'Mara MT. 2014. Sources of tooth wear variation early in life among known-aged wild ring-tailed lemurs (*Lemur catta*) at the Beza Mahafaly Special Reserve, Madagascar. *Am J Primatol* 76:1037–48.
- Darwin C. 1871. *The descent of man and selection in relation to sex*. London (UK): John Murray.
- Darwin C. 1875. *The variation of animals and plants under domestication*. London (UK): John Murray.
- Dayan T, Simberloff D, Tchernov E, Yom-Tov Y. 1989. Inter- and intraspecific character displacement in mustelids. *Ecology* 70:1526–39.
- Denes BJ, Bolton C, Illsley CS, Kok WL, Walker JV, Poetsch A, Tredwin C, Kiliaridis S, Hu B. 2019. Notch coordinates periodontal ligament maturation through regulating Lamin A. *J Dent Res* 98:1357–66.
- Dhouailly D, Godefroit P, Martin T, Nonchev S, Caraguel F, Oftedal O. 2019. Getting to the root of scales, feather and hair: as deep as odontodes?. *Exp Dermatol* 28:503–8.
- Didier DA, Stahl BJ, Zangerl R. 1994. Development and growth of compound tooth plates in *Callorhinchus milii* (Chondrichthyes, Holocephali). *J Morphol* 222:73–89.
- Di-Poï N, Milinkovitch MC. 2016. The anatomical placode in reptile scale morphogenesis indicates shared ancestry among skin appendages in amniotes. *Sci Adv* 2:e1600708.
- Dumont ER, Dávalos LM, Goldberg A, Santana SE, Rex K, Voigt CC. 2012. Morphological innovation, diversification and invasion of a new adaptive zone. *Proc Biol Sci* 279:1797–805.
- Eastman JT. 1977. The pharyngeal bones and teeth of catoctomid fishes. *Am Mid Nat* 97:68–88.
- Ellis NA, Glazer AM, Donde NN, Cleves PA, Agoglia RM, Miller CT. 2015. Distinct developmental genetic mechanisms underlie convergently evolved tooth gain in sticklebacks. *Development* 142:2442–51.
- Evans AR, Sanson GD. 2006. Spatial and functional modeling of carnivore and insectivore molariform teeth. *J Morphol* 267:649–62.
- Evans AR. 2013. Shape descriptors as ecometrics in dental ecology. *Hystrix Ital J Mamm* 24:133–40.
- Harjunmaa E, Seidel K, Häkkinen T, Renvoisé E, Corfe IJ, Kallonen A, Zhang Z-Q, Evans AR, Mikkola ML, Salazar-Ciudad I, et al. 2014. Replaying evolutionary transitions from the dental fossil record. *Nature* 512:44–8.
- Fay FH. 1982. Ecology and biology of the Pacific Walrus, *Odobenus rosmarus divergens* Illiger. *North Am Fauna* 74:1–279.
- Franchini P, Xiong P, Fruciano C, Schneider RF, Woltering JM, Hulsey CD, Meyer A. 2019. MicroRNA gene regulation in the extremely young and parallel adaptive radiations of Nicaraguan crater lake cichlid fish. *Mol Biol Evol* 36:2498–511.
- Fraser GJ, Hulsey CD, Bloomquist RF, Uyesugi K, Manley NR, Streelman JT. 2009. An ancient gene network is co-opted for teeth on old and new jaws. *PLoS Biol* 7:e100003.
- Fraser GJ, Graham A, Smith MM. 2004. Conserved deployment of genes during odontogenesis across osteichthyans. *Proc R Soc Biol* 271:2311–7.
- Fraser GJ, Cerny R, Soukup V, Bronner-Fraser M, Streelman JT. 2010. The odontode explosion: the origin of tooth-like structures in vertebrates. *Bioessays* 32:808–17.
- Fraser GJ, Smith MM. 2011. Evolution of developmental pattern for vertebrate dentitions: an oro-pharyngeal specific mechanism. *J Exp Zool B* 316B: 99–112.
- Fraser GJ, Britz R, Hall A, Johanson Z, Smith MM. 2012. Replacing the first-generation dentition in pufferfish with a unique beak. *Proc Natl Acad Sci U S A* 109:8179–84.
- Fraser GJ, Bloomquist RF, Streelman JT. 2013. Common developmental pathways link tooth shape to regeneration. *Dev Biol* 377:399–414.
- Fraser GJ, Hamed SS, Martin KJ, Hunter KD. 2019. Shark tooth regeneration reveals common stem cell characters in both human rested lamina and ameloblastoma. *Sci Rep* 9:15956.
- Freeman PW. 1988. Frugivorous and animalivorous bats (Microchiroptera): dental and cranial adaptations. *Biol J Linn Soc* 33:249–72.
- Freeman PW. 1992. Canine teeth of bats (Microchiroptera): size, shape and role in crack propagation. *Biol J Linn Soc* 45:97–115.
- Freeman PW, Lemen CA. 2007. The trade-off between tooth strength and tooth penetration: predicting optimal shape of canine teeth. *J Zool* 273:273–80.
- Gaengler P. 2000. Evolution of tooth attachment in lower vertebrates to tetrapods. In: Teaford M, Meredith Smith M, Ferguson M, editors. *Development, function and evolution of teeth*. Cambridge: Cambridge University Press. p. 173–85.
- Gallo ND, Levin LA. 2016. Fish ecology and evolution in the world's oxygen minimum zones and implications of ocean deoxygenation. *Adv Mar Biol* 74:117–98.
- Geerinckx T, De Kegel B. 2014. Functional and evolutionary anatomy of the African suckermouth catfishes (Siluriformes: Mochokidae): convergent evolution in Afrotropical and Neotropical faunas. *J Anat* 225:197–208.
- Giannini NP, Simmons NB. 2007. Element homology and the evolution of dental formulae in megachiropteran bats (Mammalia, Chiroptera, Pteropodidae). *Am Mus Novit* 3559:1–27.
- Gibbons A. 2012. An evolutionary theory of dentistry. *Science* 336:973–5.
- Gorman CE, Hulsey CD. forthcoming 2020. Non-trophic functional ecology of vertebrate teeth: a review. *Integr Comp Biol* (doi:10.1093/icb/icaa086).
- Green DR, Schulte F, Lee K-H, Pugach MK, Hardt M, Bidlack FB. 2019. Mapping the tooth enamel proteome and amelogenin phosphorylation onto mineralizing porcine tooth crowns. *Front Physiol* 10:925.

- Häärä O, Harjunmaa E, Lindfors PH, Huh SH, Fliniaux I, Åberg T, Jernvall J, Ornitz DM, Mikkola ML, Thesleff I. 2012. Ectodysplasin regulates activator-inhibitor balance in murine tooth development through Fgf20 signaling. *Development* 139:3189–99.
- Handrigan GR, Richman JM. 2010. Autocrine and paracrine Shh signaling are necessary for tooth morphogenesis, but not tooth replacement in snakes and lizards (Squamata). *Dev Biol* 337:171–86.
- Harris MP, Rohner N, Schwarz H, Perathoner S, Konstantinidis P, Nüsslein-Volhard C. 2008. Zebrafish *eda* and *edar* mutants reveal conserved and ancestral roles of ectodysplasin signaling in vertebrates. *PLoS Genet* 4:e1000206.
- Harvey PH, Kavanagh M, Clutton-Brock TH. 2009. Sexual dimorphism in primate teeth. *J Zool* 186:475–85.
- Hedrick BP, Yohe L, Vander Linden A, Davalos LM, Sears K, Sadier A, Rossiter SJ, Davies KTJ, Dumont E. 2018. Assessing soft-tissue shrinkage estimates in museum specimens imaged with diffusible iodine-based contrast-enhanced computed tomography (diceCT). *Microsc Microanal* 24:284–91.
- Heikinheimo K, Kurppa KJ, Laiho A, Peltonen S, Berdal A, Bouattour A, Ruhin B, Catón J, Thesleff I, Leivo I, et al. 2015. Early dental epithelial transcription factors distinguish ameloblastoma from keratocystic odontogenic tumor. *J Dent Res* 94:101–11.
- Hendrie CA, Brewer G. 2012. Evidence to suggest that teeth act as human ornament displays signalling mate quality. *PLoS One* 7:e42178.
- Herring SW. 1993. Functional morphology of mammalian mastication. *Am Zool* 33:289–99.
- Hoegg S, Meyer A. 2005. Hox clusters as models for vertebrate genome evolution. *Trend Genet* 21:421–4.
- Hovorakova M, Zahradnick O, Bartos M, Hurnik P, Stransky J, Stembirek J, Tucker AS. forthcoming 2020. Reawakening of ancestral dental potential as a mechanism to explain dental pathologies. *Integr Comp Biol* (doi:10.1093/icb/icaa053).
- Hulsey CD. 2006. Function of a key morphological innovation: fusion of the cichlid pharyngeal jaw. *Proc R Soc Biol* 273:669–75.
- Hulsey CD. 2009. Cichlid genomics and phenotypic diversity in a comparative context. *Integr Comp Biol* 49:618–29.
- Hulsey CD, Fraser GF, Meyer A. 2016. Biting into the genome to phenome map: developmental genetic modularity of cichlid fish dentitions. *Integr Comp Biol* 56:373–88.
- Hulsey CD, Machado-Schiaffino G, Keicher L, Ellis-Soto D, Henning F, Meyer A. 2017. The integrated genomic architecture and evolution of dental divergence in East African cichlid fishes (*Haplochromis chilotex* × *H. nyererei*). *G3* 7:3195–202.
- Hunter JP, Jernvall J. 1995. The hypocone as a key innovation in mammalian evolution. *Proc Natl Acad Sci U S A* 92:10718–22.
- Huyseune A, Sire JY. 1998. Evolution of patterns and processes in teeth and tooth-related tissues in non-mammalian vertebrates. *Eur J Oral Sci* 106:437–81.
- Huyseune A, Witten PE. 2008. An evolutionary view on tooth development and replacement in wild Atlantic salmon (*Salmo salar* L.). *Evol Dev* 10:6–14.
- Hsu YC, Li L, Fuchs E. 2014. Emerging interactions between skin stem cells and their niches. *Nat Med* 20:847–56.
- Jackman WR, Davies SH, Lyons DB, Stauder CK, Denton-Schneider BR, Jowdry A, Aigler SR, Vogel SA, Stock DW. 2013. Manipulation of *Fgf* and *Bmp* signaling in teleost fishes suggests potential pathways for the evolutionary origin of multicuspid teeth. *Evol Dev* 15:107–18.
- Jernvall J, Thesleff I. 2012. Tooth shape formation and tooth renewal: evolving with the same signals. *Development* 139:3487–97.
- Johanson Z, Underwood C, Manzanares E, Fernandez V, Clark B, Meredith-Smith M. Forthcoming 2020a. Evolution of the dentition in holocephalans (Chondrichthyes). *Integr Comp Biol*. (doi:10.1093/icb/icaa093).
- Johanson Z, Underwood C, Coates MI, Fernandez V, Clark B, Meredith Smith M. Forthcoming 2020b. The stem-holocephalan *Helodus* (Chondrichthyes; Holocephali) and the evolution of modern chimaeroid dentitions. In: Janvier P, Pradel A, editors. *Ancient fishes and their living relatives: a tribute to John G Maisey*.
- Kajiura S, Tricas T. 1996. Seasonal dynamics of dental dimorphism in the Atlantic stingray *Dasyatis sabina*. *J Exp Biol* 199:2297–306.
- Kandyba E, Leung Y, Chen YB, Widelitz R, Chuong CM, Kobiela K. 2013. Competitive balance of intrabulge BMP/Wnt signaling reveals a robust gene network ruling stem cell homeostasis and cyclic activation. *Proc Natl Acad Sci U S A* 110:1351–6.
- Karagic N, Schneider RF, Meyer A, Hulsey CD. Forthcoming 2020a. A genomic cluster containing novel and conserved genes underlies cichlid fish dental developmental convergence. *Mol Biol Evol*.
- Karagic N, Meyer A, Hulsey CD. forthcoming 2020b. Phenotypic plasticity in vertebrate dentition. *Integr Comp Biol* (doi:10.1093/icb/icaa077).
- Katsikaros K, Shine R. 1997. Sexual dimorphism in the tusked frog, *Adelotus brevis* (Anura: Myobatrachidae): the roles of natural and sexual selection. *Biol J Linn Soc* 60:39–51.
- Kawasaki K. 2009. The SCPP gene repertoire in bony vertebrates and graded differences in mineralized tissues. *Dev Genes Evol* 219:147–57.
- Kawasaki K, Suzuki T, Weiss KM. 2004. Genetic basis for the evolution of vertebrate mineralized tissue. *Proc Natl Acad Sci U S A* 101:11356–61.
- Kawasaki K, Weiss KM. 2008. SCPP gene evolution and the dental mineralization continuum. *J Dent Res* 87:520–31.
- Kawasaki K, Suzuki T, Weiss KM. 2005. Phenogenetic drift in evolution: the changing genetic basis of vertebrate teeth. *Proc Natl Acad Sci U S A* 102:18063–8.
- Keene HJ. 1991. On heterochrony in heterodonty: a review of some problems in tooth morphogenesis and evolution. *Am J Phys Anthropol* 34:251–82.
- Kemp A. 1984. A comparison of the developing dentition of *Neoceratodus forsteri* and *Callorhynchus milii*. *Proc Linn Soc NS W* 107:245–62.
- Kobiela K, Stokes N, de la Cruz J, Polak L, Fuchs E. 2007. Loss of a quiescent niche but not follicle stem cells in the absence of bone morphogenetic protein signaling. *Proc Natl Acad Sci U S A* 104:10063–8.
- Kondo S, Kuwahara Y, Kondo M, Naruse K, Mitani H, Wakamatsu Y, Ozato K, Asakawa S, Shimizu N, Shima A. 2001. The medaka rs-3 locus required for scale development encodes ectodysplasin-A receptor. *Curr Biol* 11:1202–6.
- Kouvaris K, Clune J, Kounios L, Brede M, Watson RA. 2017. How evolution learns to generalise: using the principles of

- learning theory to understand the evolution of developmental organisation. *PLoS Comput Biol* 13:e1005358.
- Kratochwil CF, Liang Y, Gerwin J, Urban S, Henning F, Machado-Schiaffino G, Woltering JM, Hulsey CD, Meyer A. 2018. Agouti related peptide 2 facilitates convergent evolution of stripe patterns across cichlid fish radiations. *Science* 362:457–60.
- Kubota T, Uyeno T. 1970. Food habits of lancetfish *Alepisaurus ferox* (order Myctophiformes) in Suruga Bay, Japan. *Jpn J Ichthyol* 17:22–8.
- Lacquaniti F, Ivanenko YP, d'Avella A, Zelik KE, Zago M. 2013. Evolutionary and developmental modules. *Front Comput Neurosci* 7:61.
- Landova M, Zahradnický O, Dumková J, Dosedelova H, Krivanek J, Hampl M, Kavkova M, Zikmund T, Gregorovicova M, Sedmera D, et al. 2020. Developmental mechanisms driving complex tooth shape in reptiles. *Dev Dyn* 249:441–64.
- Liem KF. 1973. Evolutionary strategies and morphological innovations: cichlid pharyngeal jaws. *Syst Zool* 22:425–41.
- Lipson H, Pollack JB, Suh NP. 2002. On the origin of modular variation. *Evolution* 56:1549–56.
- Martin KJ, Rasch LJ, Cooper RL, Metscher BD, Johanson Z, Fraser GJ. 2016. Sox2+ progenitors in sharks link taste development with the evolution of regenerative teeth from denticles. *Proc Natl Acad Sci U S A* 113:14769–74.
- McCormack SW, Witzel U, Watson PJ, Fagan MJ, Gröning F. 2014. The biomechanical function of periodontal ligament fibres in orthodontic tooth movement. *PLoS One* 9:e102387.
- McIntosh AF, Cox PG. 2019. The impact of digging on the evolution of the rodent mandible. *J Morphol* 280:176–83.
- McLysaght A, Guerzoni D. 2015. New genes from non-coding sequence: the role of de novo protein-coding genes in eukaryotic evolutionary innovation. *Philos Trans R Soc Lond B Biol Sci* 370:20140332.
- Mehta RS, Wainwright PC. 2007. Raptorial jaws in the throat help moray eels swallow large prey. *Nature* 449:79–82.
- Mihalitsis M, Bellwood D. 2019. Functional implications of dentition-based morphotypes in piscivorous fishes. *R Soc Open Sci* 6:190040.
- Mindell DP, Meyer A. 2001. Homology evolving. *Trends Ecol Evol* 16:434–40.
- Mitsiadis TA, Hirsinger E, Lendahl U, Goridis C. 1998. Delta-notch signaling in odontogenesis: correlation with cytodifferentiation and evidence for feedback regulation. *Dev Biol* 204:420–31.
- Moy-Thomas JA. 1939. The structure and affinities of the fossil elasmobranch fishes from the Lower Carboniferous Rocks of Glencartholm, Eskdale. *Biol Rev* 14:1–26.
- Nesse RM, Stearns SC, Omenn GS. 2006. Medicine needs evolution. *Science* 311:1071.
- Nweeia MT, Eichmiller FC, Hauschka PV, Donahue GA, Orr JR, Ferguson SH, Watt CA, Mead JG, Potter CW, Dietz R, et al. 2014. Sensory ability in the narwhal tooth organ system. *Anat Rec* 297:599–617.
- O'Connell DJ, Ho JWK, Mammoto T, Turbe-Doan A, O'Connell JT, Haseley PS, Koo S, Kamiya N, Ingber DE, Park PJ, et al. 2012. A Wnt-Bmp feedback circuit controls intertissue signaling dynamics in tooth organogenesis. *Sci Signal* 5:ra4.
- Ohno S. 1970. Evolution by gene duplication. New York (NY): Springer Science & Business Media.
- Olson E. 2017. The evolution of fangs across ray-finned fishes (Actinopterygii). St. Cloud State University, Culminating Projects in Biology. p. 22.
- Ørving T. 1967. Phylogeny of tooth tissues: evolution of some calcified tissues in early vertebrates. In: Miles AEW, editor. Structural and chemical organization of teeth Vol. 1. New York (NY): Academic Press. p. 45–110.
- Owen R. 1845. Odontography; or, a treatise on the comparative anatomy of the teeth; their physiological relations, mode of development, and microscopic structure, in the vertebrate animals. Hippolyte Baillière, London, 766 pp.
- Ovsyanikov NG. 1995. Polar bear predation upon walrus on Wrangel Island. *Bull MOIP Sec Biol* 100:3–15.
- Parmentier E, Diogo R, Fine ML. 2017. Multiple exaptations leading to fish sound production. *Fish Fisheries* 18:958–966.
- Patterson C. 1965. The phylogeny of the chimaeroids. *Phil Trans R Soc Lond B Biol Sci* 249:101–218.
- Perrin WF, Thieleking JL, Walker WA, Archer FI, Robertson KM. 2011. Common bottlenose dolphins (*Tursiops truncatus*) in California waters: cranial differentiation of coastal and offshore ecotypes. *Mar Mam Sci* 27:769–92.
- Pispa J, Thesleff I. 2003. Mechanisms of ectodermal organogenesis. *Dev Biol* 262:195–205.
- Popa EM, Anthwal N, Tucker AS. 2016. Complex patterns of tooth replacement revealed in the fruit bat (*Eidolon helvum*). *J Anat* 229:847–56.
- Popa EM, Buchtova M, Tucker AS. 2019. Revitalising the rudimentary replacement dentition in the mouse. *Development* 146:dev171363.
- Purnell MA, Bell MA, Baines DC, Hart PJB, Travis MP. 2007. Correlated evolution and dietary change in fossil stickleback. *Science* 317:1887.
- Ramsay JB, Wilga CD. 2007. Morphology and mechanics of the teeth and jaws of white-spotted bamboo sharks (*Chiloscyllium plagiosum*). *J Morphol* 268:664–82.
- Randau M, Carbone C, Turvey ST. 2013. Canine evolution in sabretoothed carnivores: natural selection or sexual selection? *PLoS One* 8:e72868.
- Rasch LJ, Martin KJ, Cooper RL, Metscher BD, Underwood CJ, Fraser GJ. 2016. An ancient dental gene set governs development and continuous regeneration of teeth in sharks. *Dev Biol* 415:347–70.
- Rose KD, Walker A, Jacobs LL. 1981. Function of the mandibular tooth comb in living and extinct mammals. *Nature* 289:583–5.
- Sadier A, Viriot L, Pantalacci S, Laudet V. 2014. The ectodysplasin pathway: from diseases to adaptations. *Trends Genet* 30:24–31.
- Salazar-Ciudad I. 2012. Tooth patterning and evolution. *Curr Opin Genet Dev* 22:585–92.
- Salazar-Ciudad I, Jernvall J. 2010. A computational model of teeth and the developmental origins of morphological variation. *Nature* 464:583–6.

- Santana SE, Cheung E. 2016. Go big or go fish: morphological specializations in carnivorous bats. *Proc Biol Sci* 283:20160615.
- Santana SE, Dumont ER. 2009. Connecting behaviour and performance: the evolution of biting behaviour and bite performance in bats. *J Evol Biol* 22:2131–45.
- Santana SE, Strait S, Dumont ER. 2011a. The better to eat you with: functional correlates of tooth structure in bats. *Funct Ecol* 25:839–47.
- Santana SE, Geipel I, Dumont ER, Kalka MB, Kalko EK. 2011b. All you can eat: high performance capacity and plasticity in the common big-eared bat, *Micronycteris microtis* (Chiroptera: Phyllostomidae). *PLoS One* 6:e28584.
- Salomies L, Eymann J, Khan I, Di-Poï N. 2019. The alternative regenerative strategy of bearded dragon unveils the key processes underlying vertebrate tooth renewal. *Elife* 8:e47702.
- Schauinsland H. 1903. Beiträge zur Entwicklungsgeschichte und Anatomie der Wirbeltiere Teil I. *Sphenodon, Callorhynchus, Chamaeleo*. *Zoologica (Stuttgart)* 39:1–98.
- Schneider RF, Meyer A. 2017. How plasticity, genetic assimilation and cryptic genetic variation may contribute to adaptive radiations. *Mol Ecol* 26:330–50.
- Seidel K, Marangoni P, Tang C, Houshmand B, Du W, Maas RL, Murray S, Oldham MC, Klein OD. 2017. Resolving stem and progenitor cells in the adult mouse incisor through gene co-expression analysis. *Elife* 6:e24712.
- Sharir A, Marangoni P, Zilionis R, Wan M, Wald T, Hu JK, Kawaguchi K, Castillo-Azofeifa D, Epstein L, Harrington K, et al. 2019. A large pool of actively cycling progenitors orchestrates self-renewal and injury repair of an ectodermal appendage. *Nat Cell Biol* 21:1102–12.
- Sharpe PT. 2001. Fish scale development: hair today, teeth and scales yesterday? *Curr Biol* 11:R751–2.
- Silverman HB, Dunbar MJ. 1980. Aggressive tusk use by the narwhal (*Monodon monoceros* L.). *Nature* 284:57–8.
- Smith MM, Coates MI. 1998. Evolutionary origins of the vertebrate dentition: phylogenetic patterns and developmental evolution. *Eur J Oral Sci* 106: 482–500.
- Smith MM. 2003. Vertebrate dentitions at the origin of jaws: when and how pattern evolved. *Evol Dev* 5:394–413.
- Smith MM, Fraser GJ, Mitsiadis T. 2009. Dental lamina as source of odontogenic stem cells: evolutionary origins and developmental control of tooth generation in gnathostomes. *J Exp Zool B Mol Dev Evol* 312B:260–80.
- Smith MM, Underwood C, Goral T, Healy C, Johanson Z. 2019. Growth and mineralogy in dental plates of the holocephalan *Harriotta raleighana* (Chondrichthyes): novel dentine and conserved patterning combine to create a unique chondrichthyan dentition. *Zool Lett* 5:11.
- Smits PD, Evans AR. 2012. Functional constraints on tooth morphology in carnivorous mammals. *BMC Evol Biol* 12:146.
- Stahl BJ. 1999. Chondrichthyes III: holocephali. In: Schultze HP, editor. *Handbook of paleoichthyology*, Vol. 4. Munich: Verlag Dr. Friedrich Pfeil. p. 1–164.
- Streelman JT, Webb JF, Albertson RC, Kocher TD. 2003. The cusp of evolution and development: a model of cichlid tooth shape diversity. *Evol Dev* 5:600–8.
- Teeling EC, Vernes SC, Dávalos LM, Ray DA, Gilbert MTP, Myers E; Bat1K Consortium. 2018. Bat biology, genomes, and the Bat1K Project: to generate chromosome-level genomes for all living bat species. *Annu Rev Anim Biosci* 6:23–46.
- Thesleff I, Sharpe P. 1997. Signaling networks regulating dental development. *Mech Dev* 67:111–23.
- Thesleff I. 2006. The genetic basis of tooth development and dental defects. *Am J Med Genet A* 140A:2530–5.
- Thiery A, Shono T, Kurokawa D, Britz R, Johanson Z, Fraser GJ. 2017. Spatially restricted dental regeneration drives pufferfish beak development. *Proc Natl Acad Sci U S A* 114:E4425–34.
- Tucker A, Sharpe P. 2004. The cutting-edge of mammalian development; how the embryo makes teeth. *Nat Rev Genet* 5:499–508.
- Tucker AS, Fraser GJ. 2014. Evolution and developmental diversity of tooth regeneration. *Semin Cell Dev Biol* 25–26:71–80.
- Uller T, Moczek AP, Watson RA, Brakefield PM, Laland KN. 2018. Developmental bias and evolution: a regulatory network perspective. *Genetics* 209:949–66.
- Underwood CJ, Johanson Z, Welten M, Metscher B, Rasch LJ, Fraser GJ, Smith MM. 2015. Development and evolution of dentition pattern and tooth order in the skates and rays (Batoidea; Chondrichthyes). *PLoS One* 10:e0122553.
- Underwood CJ, Johanson Z, Smith MM. 2016. Cutting blade dentitions in squaliform sharks form by modification of inherited alternate tooth ordering patterns. *R Soc Open Sci* 3:160385.
- Usui K, Tokita M. 2018. Creating diversity in mammalian facial morphology: a review of potential developmental mechanisms. *Evo Dev* 9:15.
- Van de Peer Y, Taylor JS, Meyer A. 2003. Are all fishes ancient polyploids?. *J Struct Funct Genome* 3:65–73.
- Vonk FJ, Admiraal JF, Jackson K, Reshef R, de Bakker MAG, Vanderschoot K, van den Berge I, van Atten M, Burgerhout E, Beck A, et al. 2008. Evolutionary origin and development of snake fangs. *Nature* 454:630–3.
- Wang XP, Suomalainen M, Felszeghy S, Zelarayan LC, Alonso MT, Plikus MV, Maas RL, Chuong CM, Schimmang T, Thesleff I. 2007. An integrated gene regulatory network controls stem cell proliferation in teeth. *PLoS Biol* 5:e159.
- Whitenack LB, Motta PJ. 2010. Performance of shark teeth during puncture and draw: implications for the mechanics of cutting. *Biol J Linn Soc Lond* 100:271–86.
- Whitlock JA, Richman JM. 2013. Biology of tooth replacement in amniotes. *Int J Oral Sci* 5:66–70.
- Witten PE, Huyseune A. 2009. A comparative view on mechanisms and functions of skeletal remodelling in teleost fish, with special emphasis on osteoclasts and their function. *Biol Rev* 84:315–46.
- Wu DD, Irwin DM, Zhang YP. 2011. *De novo* origin of human protein-coding genes. *PLoS Genet* 7:e1002379.
- Wu DD, Zhang YP. 2013. Evolution and function of *de novo* originated genes. *Mol Phylogenet Evol* 67:541–5.
- Wu P, Wu X, Jiang TX, Elsey RM, Temple BL, Divers SJ, Glenn TC, Yuan K, Chen MH, Widelitz RB, et al. 2013.

- Specialized stem cell niche enables repetitive renewal of alligator teeth. *Proc Natl Acad Sci U S A* 110:E2009–18.
- Wu P, Zhang Y, Xing Y, Xu W, Guo H, Deng F, Ma X, Li Y. 2019. The balance of *Bmp6* and *Wnt10b* regulates the telogen–anagen transition of hair follicles. *Cell Commun Signal* 17:16.
- Xiong P, Schneider RF, Hulsey CD, Meyer A, Franchini P. 2019. Conservation and novelty in the microRNA genomic landscape of hyperdiverse cichlid fishes. *Sci Rep* 9:13848.
- Xiong P, Hulsey CD, Meyer A, Franchini P. 2018. Evolutionary divergence of 3'-UTRs in cichlid fishes. *BMC Genome* 19:433.